

Normal Cell Differentiation Potential of Cancer Stem Cells Without Reprogramming Pluripotent Factors: a Novel Strategy in Stem Cell-Based Therapy for Tissue Regeneration

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Abstract:

Stem cells hold great promise for tissue regeneration and have the potential to treat many incurable degenerative diseases. Cancer stem cells (CSCs), or cancer initiating cells, have the ability to self-renew and differentiate into heterogeneous lineages of cancer cells. Current stem cell therapies face limitations, such as limited stem cell sources, time consumption, tumor formation, and immune rejection upon allogeneic transplantation. Allogeneic stem cell treatments simplify stem cell manufacturing and reduce transplant time, but their therapeutic potential is limited by human leukocyte antigen (HLA)-matched donors. CSCs retain characteristics essential for tissue regeneration. However, several limitations hinder cancer stem cell reprogramming with pluripotent factors. The development of 3D culture models for tissue imitating extracellular matrix in cancer cell lines aims to enhance CSC enrichment. This mini-review focuses on a new strategy for treating incurable degenerative diseases involving *in vitro* and *in vivo* 3D cancer models and the induced differentiation of CSCs into mature normal cell types. This allows tissue survival without immune rejection and offers a safe alternative to cancer stem cell reprogramming with pluripotent factors. In conclusion, preservation and banking of allogeneic CSCs offer an alternative, readily available, and safe strategy that can be used to facilitate stem cell-based cell therapy.

Keywords — Stem cell therapy, Cancer stem cells, iPSCs, Stem cell bank, Allogeneic transplantation, Differentiation therapy.

I. INTRODUCTION

Cancer stem cells (CSCs), or cancer initiating cells, are a small population of cancer cells essential for tumor initiation, maintenance, metastasis, and recurrence, as well as having the ability to self-renew, proliferate, and differentiate into "heterogeneous lineages" of cancer cells [1-10]. CSCs are responsible for the chemoresistance, recurrence, and metastasis of tumors [11-13]. CSCs were first identified in human acute myeloid leukemia [14] and have also been identified in many other human solid

tumors, such as breast [15], brain [16], ovarian [17], colon [18], skin [19], prostate [20], pancreatic [21], liver [22], and lung carcinomas [23], etc. CSCs form "niches" in the tumor microenvironment (TME), which mainly contains fibroblasts, immune cells, mesenchymal cells, endothelial cells, and extracellular matrix [24-27]. The niche is required for their survival and controls the self-renewal and differentiation of CSCs, similar to normal stem cells [28-30]. CSCs modulate their TME to escape immune detection through different mechanisms [31], and their ability to differentiate into heterogeneous lineages of cancer cells has been

demonstrated by many experimental models [32, 33]. CSCs avoid elimination by natural killer (NK) cells and cytotoxic T cells through various immune modulation mechanisms [34-36]. The CSCs of many tumors resist NK cell killing due to an “immunosuppressive” microenvironment [37, 38]. The tumor microenvironment weakens cytotoxic T cell function, reducing “immunogenicity” [39-41]. Treating cancers through the induction of differentiation has been an attractive and practical alternative to killing them through cytotoxicity [42-44]. However, the mechanisms responsible for differentiation vary among different tumor types [45-47]. Moreover, clinical studies reveal that CSCs are less tumorigenic or not tumorigenic at all after differentiation [48].

Stem cell therapy is a feasible clinical practice for the treatment of many “incurable degenerative diseases” [49], such as hematopoietic diseases [50], diabetes [51], cancer [52], eye diseases [53], stroke [54], neurodegenerative disorders [55], joint diseases [56], cardiovascular diseases [57], autoimmune diseases [58], and blistering skin diseases [59], etc. Based on stem cells' capacity to divide indefinitely and their functional differentiation into any type of tissue, the discovery of stem cells has offered a potential treatment for many incurable degenerative diseases, restoring organ function [60-64]. However, current stem cell therapies face limitations such as limited stem cell sources, extreme time consumption for autologous cell therapy, immune rejection upon allogeneic transplantation, ethical issues concerning embryonic stem cell (ESC) application, tumor formation, and "epigenetic defects," necessitating new strategies [65-72]. Allogeneic induced pluripotent stem cells (iPSCs) have been used for disease treatment, but their prolonged somatic cell reprogramming and tumor formation make them challenging for stem cell-based therapies [73, 74]. Furthermore, extended use of immunosuppressive medications in allogeneic iPSC therapy has a greater tendency to cause many side effects [75, 76].

Tissue regeneration gives rise to new tissues in order to restore damaged tissues' function [77-79]. CSCs, similar to iPSCs, retain characteristics essential for tissue regeneration such as high capability for self-renewal, “pluripotency,” differentiation potential, and a high level of

proliferation [80-88]. Interestingly, new connections have been discovered between iPSC generation and tumor cell plasticity [89]. However, the low efficiency of cancer cell reprogramming with pluripotent factors limits the use of “cancer-iPSCs” in stem cell-based therapies [90, 91].

Extracellular matrix (ECM) provides a convenient environment that supports cell expansion and tissue formation [92, 93]. 3D scaffolds enable rapid tissue repair using an artificial ECM environment [94, 95]. 3D culture methods vary based on whether they include scaffolds or not [96]. Effective 3D cell scaffold methods for enriching CSCs are needed due to limitations in existing methods [97].

The unique properties of CSCs, including self-renewal, pluripotency, immune modulation, and the ability to differentiate into heterogeneous lineages of cancer cells, offer novel strategies for the application of allogeneic CSCs to a large population for disease modeling, drug discovery, and the revolution of economically viable stem cell-based therapy, which could significantly reduce immune rejection. However, further clinical studies are required to ensure efficient effects *in vitro* and *in vivo*. In fact, normalization of CSC differentiation can be activated in the future to treat multiple incurable degenerative diseases.

II. CANCER STEM CELLS ARE QUALIFIED STEM CELL CANDIDATES FOR TISSUE REGENERATION

Tissue regeneration gives rise to new tissues in order to restore damaged tissues' function [98, 99]. The high proliferative and differentiation capacities of stem cells are crucial for treating various incurable degenerative diseases and injuries through differentiation into mature cells [100-102]. Scientists are investigating novel ways to improve stem cell therapy for tissue regeneration [103]. Tissue regeneration and “tumorigenesis” have common characteristics that differ slightly, whereas tissue regeneration is misused by cancer cells [104].

Self-renewal involves stem cells dividing to produce daughter cells with identical prospects of development, enabling excellent replication while maintaining vast developmental and replicative capacity [105]. Stem cell self-renewal is crucial for effective tissue regeneration [106]. Stem cells

replicate and give rise to duplicate cells of the original cells [107]. The self-renewal and differentiation of stem cells are determined by signals received from stem cell niches [108-110]. Similar to normal stem cells, the CSC niche governs the self-renewal and differentiation properties of CSCs [111]. Optimized self-renewal in cancer stem cells is crucial for effective cell development, similar to tissue regeneration [112].

Many tumor CSCs retain pluripotency, the ability of individual cells to differentiate into all cell types of the adult human body, regulated by the core transcriptional factors octamer-binding transcription factor 4 (OCT4), sex-determining region Y-box 2 (SOX2), and homeobox protein NANOG (NANOG) [113-117]. Human pluripotent stem cells, such as ESCs and iPSCs, can differentiate into all cell types, providing unprecedented opportunities for cell therapies for the treatment of incurable degenerative diseases and injuries [118, 119]. ESCs harvested from pre-implantation embryos have the potential to differentiate into any cell type derived from the three germ layers of ectoderm, mesoderm, and endoderm [120-122]. Furthermore, iPSCs can be generated from various differentiated cell types through the expression of a set of defined transcription factors [123]. In 2006, Takahashi and Yamanaka discovered that somatic cells could be reprogrammed into a pluripotent state involving four transcription factors, OCT4, SOX2, c-MYC, and Krüppel-like factor 4 (KLF4), and that these reprogrammed iPSCs were similar to human ESCs [124-126]. Tumorigenesis exhibit similarities between embryonic development and the ability of ESCs, and CSCs to differentiate into heterogeneous lineages of cells [127]. Cancer maturation involves proliferation through transformed protein expression and signaling pathways, promoting the survival and proliferation of premature CSCs [128-131]. Thus, pluripotent stem cells such as ESCs, iPSCs, and CSCs share similarities between self-renewal, tumor formation, rapid proliferation, and cellular plasticity [132-135].

Stem cell plasticity involves cells' ability to flexibly change their characteristics, which is essential for tissue regeneration and the differentiation of stem cells, whereas CSC plasticity is regulated by tumor microenvironmental signals,

playing a crucial role in therapeutic resistance, tumor relapse, and metastasis [136-141]. Hence, modulating cell plasticity in order to obtain differentiated cells of interest has caught the attention of scientists [142, 143].

Epithelial-to-mesenchymal transition (EMT) is a genetic process involving epithelial cells transforming into mesenchymal phenotypes, impacting embryonic development, tissue regeneration, tumor progression, and therapy resistance, resulting in invasion and metastasis of tumors [144-147]. iPSC generation from somatic cells involves mesenchymal-to-epithelial transition, the reverted process of EMT [148, 149]. Evidence suggests cancer cells retain tumor-initiating potential due to their plasticity modulation, which supports EMT mechanisms [150]. Clinical studies show that EMT is associated with tissue regeneration in several tissue models [151, 152]. Hence, there is a tangential connection between tissue regeneration and tumorigenesis [153].

Ensuring cells thrive in a conducive environment with cells, scaffolds, and growth factors for proliferation and differentiation is vital to tissue regeneration [154-158]. Repair of injured tissue involves the production of extracellular matrix components, which are restored over time to mimic normal tissue, modulating the cellular processes for tissue reconstruction [159]. ECM signals cells, regulating proliferation, migration, and differentiation; thus, tissue formation and regeneration heavily rely on cellular interaction with ECM [160]. Extracellular matrix scaffolds promote tissue-specific remodeling and repair in various organs, fostering a regenerative microenvironment and functional reconstruction [161-163]. 3D culture methods vary based on whether they include scaffolds or not [164, 165]. 3D scaffolds enable rapid tissue repair using an artificial ECM environment [166].

A precise *in vitro* and *in vivo* demonstration of tissue of interest is under development based on 3D culture models in order to imitate extracellular matrix and provide an appropriate niche for CSC enrichment in various cancer cell lines, such as cholangiocarcinoma [167], lung carcinoma [168], colorectal cancer [169], acute myeloid leukemia [170], glioblastoma [171], hepatocellular carcinoma

[172], melanoma [173], breast [174], prostate [175], neuroblastoma [176], ovarian cancer [177], etc. 3D culture models are more favorable than two-dimensional culture models, but due to variations in "biomaterials," "manufacturing methods," and tumor heterogeneity, the development of a common 3D culture model demonstrating all tumor niches is uncertain [178-180]. And also, it is crucial to advance current knowledge about various tumor differentiation mechanisms [181, 182].

III. NORMAL CELL DIFFERENTIATION POTENTIAL OF CANCER STEM CELLS

Differentiation therapy offers promising cancer treatment for malignant cells with tumorigenicity reduction rather than cytotoxic lysis [183]. CSCs are naturally capable of differentiation into heterogenous lineages of tumor cells and progress through a decreased ability to differentiate into a normal cell state [184].

Differentiation therapy induces cancer cells to differentiate into benign or normal cells [185, 186]. CSC differentiation therapy involves tumorigenic CSC differentiation into low-tumorigenic stem cells or mature cells, reducing the "CSC pool" for cancer eradication [187, 188].

Solid tumor differentiation is induced by differentiation inducers both *in vitro* and *in vivo* [189]. Currently, leukemia treatment is mainly focused on leukemic cell differentiation with various differentiation inducers [190]. All trans-retinoic acid induces differentiation, which transforms acute promyelocytic leukemia into mature granulocytes [191]. Melanoma cells have the capacity to differentiate into "mesenchymal lineages" [192]. Differentiation therapy for liver cancer involves altering hepatocyte dedifferentiation and promoting tumor differentiation into normal liver cells [193]. Redifferentiation of nodule hepatocytes gives rise to normal liver characteristics [194]. Neuroblastoma cells differentiate into normal adult neuronal cells under specific growth conditions [195-197]. Interleukin-15 gives rise to normal epithelial differentiation of renal CSCs *in vitro* [198]. Induced differentiation of osteosarcoma-initiating cells gives rise to adipocytes, restraining tumorigenicity [199]. All trans-retinoic acid promotes osteogenic

differentiation in osteosarcoma cells [200]. Induced differentiation of hepatocellular carcinoma cells generates normal hepatocyte-like cells [201]. Breast cancer cells undergo transdifferentiation, generating adipocytes both *in vitro* and *in vivo* [202]. Polyphenols, derived from plants, have potential therapeutic applications in anti-cancer therapies due to their reduced side effects and antioxidant properties, as well as their targeting of signaling pathways regulating cellular processes such as proliferation, apoptosis, and differentiation [203, 204].

Thus, CSCs have the potential to produce new tissues upon induced differentiation without reprogramming pluripotent factors *in vitro* or *in vivo*, which in turn promotes tissue regeneration. Transdifferentiation and dedifferentiation of tumor cells, or redifferentiation of CSCs into normal adult cells, exhibit potential connections with tissue regeneration [205, 206]. Lacking information about stem cell niche pathways controlling cellular quiescence and self-renewal in normal stem cells and CSCs is considered a major hurdle that could potentially hinder the application of the CSC differentiation therapy concept to stem cell-based therapies [207]. Understanding the CSC differentiation mechanisms of solid tumors will advance stem cell-based therapies in the future.

IV. UNSHAKABLE IMMUNE CONTROL OF CANCER STEM CELLS

"Immune privilege" offers shelter to vital tissues against foreign antigens [208]. CSCs develop defense mechanisms against immune detection and destruction using immune modulation strategies that enable them to bypass innate and adaptive immune control [209-211]. CSC immunological functions include evasion from immune clearance, induction of "tumor-antigen-specific T cells," activation of regulatory immune cells, and release of immune suppressive molecules in the tumor microenvironment [212-215].

Classical major histocompatibility complex (MHC) genes encode glycoproteins crucial to the immune response [216-218]. Human leukocyte antigen (HLA) genes are highly polymorphic in the human genome [219, 220]. MHC molecules recognize T cell and NK cell receptors [221-224].

In many solid tumors, CSCs show low expression of MHC-I and II [225-227]. During embryonic development, ESCs either express MHC-I at a low level or have no MHC-II expression at all, making them poor targets for the mother's immune cells [228, 229]. Similarly, downregulation of MHC-I expression in various CSCs makes them resistant to immune detection, evading T cells and NK cell killing [230-232]. Many tumor CSCs maintain restricted MHC-I levels to avoid NK cell recognition [233, 234].

HLA-G is a nonclassical HLA class-I molecule expressed in placental trophoblasts and CSCs [235, 236]. Tumor cells use *HLA-G* expression to bypass immune detection in the host [237, 238]. CSCs upregulate MHC class 1 or Human Leukocyte Antigen-G (*HLA-G*) expression to inhibit NK cell activation [239, 240].

Core tumor-associated cells in TME involve neutrophils [241], macrophages [242], regulatory T cells [243], and myeloid suppressor cells [244], creating an immunosuppressive environment for growth. Tumor cells use various mechanisms to bypass NK-mediated immune destruction [245]. Tumor microenvironments downregulate natural killer group 2, member D receptor ligands, causing tumors to escape immune detection [246-248]. Limited NK cell infiltration limits tumor elimination. NK cells can also be excluded from TME [249]. Defects in dendritic cell maturation, which are crucial in immune responses, lead to tumor progression [250, 251]. Tumor-associated macrophages play a crucial role in innate immune responses and in the likelihood of tumor formation, and this immunosuppressive environment can weaken T-cells and promote tumor development using various strategies [252-255]. Myeloid-derived suppressor cell accumulation is promoted by various stress conditions that weaken immature myeloid cell differentiation, suppressing "antitumor immune responses" in order to maintain the CSC population [256-258]. Neutrophils contribute to tumor progression both directly and indirectly by influencing the tumor microenvironment and inducing tumor progression by producing cytokines, chemokines, reactive oxygen species, proteinases, and toxins that are able to alter the tumor microenvironment [259-264]. In many solid tumors,

regulatory T cells and regulatory B cells facilitate tumor development via several immunosuppressive mechanisms [265-271].

Concerning these basic immunomodulatory mechanisms, CSCs are tricky players to escape immune detection and survive within a foreign host, which is a fundamental requirement in allogeneic stem cell treatments.

V. DISCUSSION

A stem cell bank stores donor stem cells prior to clinical application [272, 273]. The generation of MHC/HLA-matched allogeneic stem cell banks for a large population could significantly reduce immunological rejection and the cost of stem cell-based cell therapy [274].

Although autologous methods offer a reduced risk of immunological rejection compared to allogeneic donor methods [275, 276], autologous transplantation is costly and requires a lengthy process, limiting its therapeutic potential [277]. There is no need to identify a HLA-matched donor in autologous transplantation, but due to several shortcomings, allogeneic stem cell banks are critical to improving their therapeutic efficacy, and it is crucial to identify a HLA-matched donor in order to prevent immune rejection upon allogeneic transplantation [278, 279]. Allogeneic treatments offer viable manufacturing of multiple "allografts" from a single donor, simplifying the stem cell manufacturing process and reducing the time to transplant stem cells to patients [75]. However, allogeneic stem cell therapy is limited to HLA-matched donors [280].

CSCs resemble iPSC characteristics, potentially allowing reprogramming in order to generate an infinite CSC pool, but cancer cell reprogramming faces many challenges due to the negative association of cancer cell diversity with successful reprogramming [281, 282]. Furthermore, cells reprogrammed with pluripotent factors have a high potential for tumor formation [312].

CSCs possess immune privilege, enabling them to bypass immune control within the host by developing immune modulation strategies against immune responses [283, 284].

Presently, cancer therapy considers eradicating cancer via differentiation of CSCs rather than killing

them through cytotoxicity [285-287]. Cancers such as leukemia [288, 289], glioma [290], liver [291], colon [292], breast [293], skin [237], ovarian [294], lung carcinoma [295], neuroblastoma [296], and melanoma [297], etc. offer positive results to differentiation therapy. For instance, successful application of polyphenols manifests differentiation in many tumors [298, 299]. Resveratrol exhibits antitumor effects in various cancers, modulating tumorigenesis [300, 301] and inducing the differentiation of glioma stem cells into non-tumorigenic cells, potentially eliminating tumors [302]. Flavonoids are plant compounds with therapeutic effects on glioma cells, increasing differentiation biomarkers and preventing cancer [303]. Chlorogenic acid induces the differentiation of neuroblastoma cells *in vitro* and *in vivo* by inhibiting acetyl-CoA acetyltransferase [304]. Kaempferol and melatonin promote neuroblastoma differentiation [305]. However, incomprehensible details on normal differentiation pathway mechanisms need to be resolved by researchers [189].

Generation of the desired CSC line of interest *in vitro* and *in vivo* is viable [306, 307]. However, isolation and *in vitro* and *in vivo* replication of CSCs are limited due to the lack of CSC model development [308, 309]. Furthermore, limitations in *in vitro* and *in vivo* two-dimensional cultures of CSC restrict CSC-based clinical studies [310, 311]. Interestingly, 3D cellular scaffolding models offer a pragmatic tumor microenvironment, which is crucial for tissue regeneration [312, 313]. 3D tumor models enable *in vitro* and *in vivo* CSC replication, enabling

CSCs to generate unlimited stem cells for stem cell-based therapy [314, 315]. Effective 3D cell scaffold methods for enriching CSCs are needed due to limitations in existing methods [316, 317]. Thus, standardized 3D tumor models could revolutionize *in vitro* and *in vivo* replication and expansion of CSCs in preclinical settings [318, 319].

VI. CONCLUSION

In conclusion, immune modulation of CSCs prevents the elimination of CSCs by the immune system, enabling allogeneic CSCs for various clinical studies. The unique properties of CSCs, including self-renewal, pluripotency, immune modulation, and the ability to differentiate into heterogeneous lineages of cancer cells, offer novel strategies for the application of MHC/HLA-matched allogeneic cancer stem cells to a large population for tissue regeneration, disease modeling, drug discovery, and the revolution of economically viable stem cell-based therapy, which could significantly reduce immune rejection. However, further clinical studies are required to ensure safe and efficient effects *in vivo*. The use of allogeneic CSC for the regeneration of incurable degenerative diseases based on differentiation therapy may be an alternative to iPSC technology. Therefore, CSC may represent a more promising stem cell candidate in stem cell-based therapy, and I believe that CSC will be one of the stem cell researchers' great interests, improving patient outcomes.

CONFLICT OF INTEREST STATEMENT

The author declares no conflict of interest.

REFERENCES

- [1] C. Li, D.G. Heidt, P. Dalerba, C.F. Burant, L. Zhang, V. Adsay, M. Wicha, M.F. Clarke, D.M. Simeone, Identification of pancreatic cancer stem cells, *Cancer research*, 67 (2007) 1030-1037.
- [2] A. Scopelliti, P. Cammareri, V. Catalano, V. Saladino, M. Todaro, G. Stassi, Therapeutic

AUTHOR CONTRIBUTIONS

JJA contributed to the conception and design of the study and wrote sections of the manuscript.

- implications of cancer initiating cells, Expert opinion on biological therapy, 9 (2009) 1005-1016.
- [3] H. Huldani, S.A. Jasim, K.N. Sergeenva, D.O. Bokov, W.K. Abdelbasset, R. Turakulov, M.E. Al-Gazally, B. Ahmadzadeh, Z.H. Jawhar, H. Siahmansouri, Mechanisms of cancer stem cells drug resistance and the pivotal role of HMGA2, Pathology-Research and Practice, 234 (2022) 153906.
- [4] R.W. Cho, M.F. Clarke, Recent advances in cancer stem cells, Current opinion in genetics & development, 18 (2008) 48-53.
- [5] N.Y. Frank, T. Schatton, M.H. Frank, The therapeutic promise of the cancer stem cell concept, The Journal of clinical investigation, 120 (2010) 41-50.
- [6] M. Maugeri-Saccà, P. Vigneri, R. De Maria, Cancer stem cells and chemosensitivity, Clinical Cancer Research, 17 (2011) 4942-4947.
- [7] Z. Huang, T. Wu, A.Y. Liu, G. Ouyang, Differentiation and transdifferentiation potentials of cancer stem cells, Oncotarget, 6 (2015) 39550.
- [8] B. Bao, Z. Wang, S. Ali, A. Ahmad, A.S. Azmi, S.H. Sarkar, S. Banerjee, D. Kong, Y. Li, S. Thakur, F.H. Sarkar, Metformin inhibits cell proliferation, migration and invasion by attenuating CSC function mediated by deregulating miRNAs in pancreatic cancer cells, Cancer Prev Res (Phila), 5 (2012) 355-364.
- [9] F. Li, B. Tiede, J. Massague, Y. Kang, Beyond tumorigenesis: cancer stem cells in metastasis, Cell Res, 17 (2007) 3-14.
- [10] C. Peitzsch, A. Tyutyunnykova, K. Pantel, A. Dubrovskaya, Cancer stem cells: The root of tumor recurrence and metastases, in: Seminars in cancer biology, Elsevier, 2017, pp. 10-24.
- [11] R. Ojha, S. Bhattacharyya, S.K. Singh, Autophagy in cancer stem cells: a potential link between chemoresistance, recurrence, and metastasis, BioResearch open access, 4 (2015) 97-108.
- [12] L.N. Abdullah, E.K.-H. Chow, Mechanisms of chemoresistance in cancer stem cells, Clinical and translational medicine, 2 (2013) 1-9.
- [13] J.A. McCubrey, S.L. Abrams, T.L. Fitzgerald, L. Cocco, A.M. Martelli, G. Montalto, M. Cervello, A. Scalisi, S. Candido, M. Libra, Roles of signaling pathways in drug resistance, cancer initiating cells and cancer progression and metastasis, Advances in biological regulation, 57 (2015) 75-101.
- [14] W.-I. Chan, B.J. Huntly, Leukemia stem cells in acute myeloid leukemia, in: Seminars in oncology, Elsevier, 2008, pp. 326-335.
- [15] M. Al-Hajj, M.S. Wicha, A. Benito-Hernandez, S.J. Morrison, M.F. Clarke, Prospective identification of tumorigenic breast cancer cells, Proceedings of the National Academy of Sciences, 100 (2003) 3983-3988.
- [16] S.G. Piccirillo, E. Binda, R. Fiocco, A.L. Vescovi, K. Shah, Brain cancer stem cells, Journal of molecular medicine, 87 (2009) 1087-1095.
- [17] M.J. Kwon, Y.K. Shin, Regulation of ovarian cancer stem cells or tumor-initiating cells, International journal of molecular sciences, 14 (2013) 6624-6648.
- [18] L. Ricci-Vitiani, E. Fabrizi, E. Palio, R. De Maria, Colon cancer stem cells, Journal of Molecular Medicine, 87 (2009) 1097-1104.
- [19] C.S. Colmont, K.G. Harding, V. Piguat, G.K. Patel, Human skin cancer stem cells: a tale of mice and men, Experimental dermatology, 21 (2012) 576-580.
- [20] J.R. Packer, N.J. Maitland, The molecular and cellular origin of human prostate cancer, Biochimica et Biophysica Acta (BBA)-Molecular Cell Research, 1863 (2016) 1238-1260.

- [21] D.M. Simeone, Pancreatic cancer stem cells: implications for the treatment of pancreatic cancer, *Clinical cancer research*, 14 (2008) 5646-5648.
- [22] T.K.W. Lee, V.C.H. Cheung, I.O.L. Ng, Liver tumor-initiating cells as a therapeutic target for hepatocellular carcinoma, *Cancer letters*, 338 (2013) 101-109.
- [23] A. Eramo, T.L. Haas, R. De Maria, Lung cancer stem cells: tools and targets to fight lung cancer, *Oncogene*, 29 (2010) 4625-4635.
- [24] J. López de Andrés, C. Griñán-Lisón, G. Jiménez, J.A. Marchal, Cancer stem cell secretome in the tumor microenvironment: a key point for an effective personalized cancer treatment, *Journal of hematology & oncology*, 13 (2020) 1-22.
- [25] M. Prieto-Vila, R.-u. Takahashi, W. Usuba, I. Kohama, T. Ochiya, Drug resistance driven by cancer stem cells and their niche, *International journal of molecular sciences*, 18 (2017) 2574.
- [26] T.S. Zhu, M.A. Costello, C.E. Talsma, C.G. Flack, J.G. Crowley, L.L. Hamm, X. He, S.L. Hervey-Jumper, J.A. Heth, K.M. Muraszko, Endothelial cells create a stem cell niche in glioblastoma by providing NOTCH ligands that nurture self-renewal of cancer stem-like cells, *Cancer research*, 71 (2011) 6061-6072.
- [27] S. Nallanthighal, J.P. Heiserman, D.-J. Cheon, The role of the extracellular matrix in cancer stemness, *Frontiers in cell and developmental biology*, 7 (2019) 86.
- [28] N. Takakura, Formation and regulation of the cancer stem cell niche, *Cancer Science*, 103 (2012) 1177-1181.
- [29] P. Nallasamy, R.K. Nimmakayala, S. Parte, A.C. Are, S.K. Batra, M.P. Ponnusamy, Tumor microenvironment enriches the stemness features: The architectural event of therapy resistance and metastasis, *Molecular cancer*, 21 (2022) 1-25.
- [30] S.-Y. Yi, Y.-B. Hao, K.-J. Nan, T.-L. Fan, Cancer stem cells niche: a target for novel cancer therapeutics, *Cancer treatment reviews*, 39 (2013) 290-296.
- [31] K. Deepak, R. Vempati, G.P. Nagaraju, V.R. Dasari, S. Nagini, D. Rao, R.R. Malla, Tumor microenvironment: Challenges and opportunities in targeting metastasis of triple negative breast cancer, *Pharmacological research*, 153 (2020) 104683.
- [32] T. Denysenko, L. Gennero, C. Juenemann, I. Morra, P. Masperi, V. Ceroni, A. Pragliola, A. Ponzetto, A. Melcarne, Heterogeneous phenotype of human glioblastoma: in vitro study, *Cell Biochemistry and Function*, 32 (2014) 164-176.
- [33] T. Borovski, L. Vermeulen, M.R. Sprick, J.P. Medema, One renegade cancer stem cell?, *Cell Cycle*, 8 (2009) 803-808.
- [34] E.E. Abd El-Fattah, H.M. Selim, Reprogramming immune microenvironment modulates CD47 cancer stem cells in hepatocellular carcinoma, *Int Immunopharmacol*, 113 (2022) 109475.
- [35] J.-M. Hsu, W. Xia, Y.-H. Hsu, L.-C. Chan, W.-H. Yu, J.-H. Cha, C.-T. Chen, H.-W. Liao, C.-W. Kuo, K.-H. Khoo, STT3-dependent PD-L1 accumulation on cancer stem cells promotes immune evasion, *Nature communications*, 9 (2018) 1908.
- [36] M. Sultan, D. Vidovic, A.S. Paine, T.T. Huynh, K.M. Coyle, M.L. Thomas, B.M. Cruickshank, C.A. Dean, D.R. Clements, Y. Kim, Epigenetic silencing of TAP1 in aldefluor+ breast cancer stem cells contributes to their enhanced immune evasion, *Stem Cells*, 36 (2018) 641-654.
- [37] C. Guillerey, NK cells in the tumor microenvironment, *Tumor Microenvironment: Hematopoietic Cells--Part B*, (2020) 69-90.
- [38] A. Groth, S. Klöss, E. Pogge von Strandmann, U. Koehl, J. Koch, Mechanisms of tumor and viral immune escape from natural killer cell-mediated surveillance, *Journal of innate immunity*, 3 (2011) 344-354.
- [39] Q. Wu, L. You, E. Nepovimova, Z. Heger, W. Wu, K. Kuca, V. Adam, Hypoxia-inducible factors:

- Master regulators of hypoxic tumor immune escape, *Journal of hematology & oncology*, 15 (2022) 1-18.
- [40] E. Quaglino, L. Conti, F. Cavallo, Breast cancer stem cell antigens as targets for immunotherapy, in: *Seminars in immunology*, Elsevier, 2020, pp. 101386.
- [41] G. Gupta, G. Merhej, S. Saravanan, H. Chen, Cancer resistance to immunotherapy: What is the role of cancer stem cells?, *Cancer Drug Resistance*, 5 (2022) 981.
- [42] M. Bizzarri, A. Giuliani, A. Cucina, M. Minini, Redifferentiation therapeutic strategies in cancer, *Drug Discovery Today*, 25 (2020) 731-738.
- [43] M. Najafi, K. Mortezaee, J. Majidpoor, Cancer stem cell (CSC) resistance drivers, *Life sciences*, 234 (2019) 116781.
- [44] M.A. Salvador, J. Wicinski, O. Cabaud, Y. Toiron, P. Finetti, E. Josselin, H. Lelievre, L. Kraus-Berthier, S. Depil, F. Bertucci, The histone deacetylase inhibitor abexinostat induces cancer stem cells differentiation in breast cancer with low Xist expression, *Clinical cancer research*, 19 (2013) 6520-6531.
- [45] P.H. Nguyen, J. Giraud, C. Staedel, L. Chambonnier, P. Dubus, E. Chevet, H. Bœuf, X. Gauthereau, B. Rousseau, M. Fevre, All-trans retinoic acid targets gastric cancer stem cells and inhibits patient-derived gastric carcinoma tumor growth, *Oncogene*, 35 (2016) 5619-5628.
- [46] X. Zhang, B. Hu, Y.F. Sun, X.W. Huang, J.W. Cheng, A. Huang, H.Y. Zeng, S.J. Qiu, Y. Cao, J. Fan, Arsenic trioxide induces differentiation of cancer stem cells in hepatocellular carcinoma through inhibition of LIF/JAK1/STAT3 and NF - kB signaling pathways synergistically, *Clinical and Translational Medicine*, 11 (2021) e335.
- [47] D. Klevebring, G. Rosin, R. Ma, J. Lindberg, K. Czene, J. Kere, I. Fredriksson, J. Bergh, J. Hartman, Sequencing of breast cancer stem cell populations indicates a dynamic conversion between differentiation states in vivo, *Breast cancer research*, 16 (2014) 1-7.
- [48] L. Vermeulen, M. Sprick, K. Kemper, G. Stassi, J. Medema, Cancer stem cells—old concepts, new insights, *Cell Death & Differentiation*, 15 (2008) 947-958.
- [49] X. Zhao, K. Cui, Z. Li, The role of biomaterials in stem cell-based regenerative medicine, *Future Medicinal Chemistry*, 11 (2019) 1777-1790.
- [50] B.M. Abdallah, M. Kassem, The use of mesenchymal (skeletal) stem cells for treatment of degenerative diseases: current status and future perspectives, *Journal of cellular physiology*, 218 (2009) 9-12.
- [51] K. Docherty, A.S. Bernardo, L. Vallier, Embryonic stem cell therapy for diabetes mellitus, in: *Seminars in cell & developmental biology*, Elsevier, 2007, pp. 827-838.
- [52] S.K. Baird, Mesenchymal stem cells: how can we realize their therapeutic potential in cancer therapy?, *Journal of Clinical & Experimental Pathology*, 5 (2015) 2161-0681.1000206.
- [53] B. Mead, M. Berry, A. Logan, R.A. Scott, W. Leadbeater, B.A. Scheven, Stem cell treatment of degenerative eye disease, *Stem cell research*, 14 (2015) 243-257.
- [54] Y.-H. Chang, K.-C. Wu, H.-J. Harn, S.-Z. Lin, D.-C. Ding, Exosomes and stem cells in degenerative disease diagnosis and therapy, *Cell transplantation*, 27 (2018) 349-363.
- [55] J.S. Lunn, S.A. Sakowski, J. Hur, E.L. Feldman, Stem cell technology for neurodegenerative diseases, *Annals of neurology*, 70 (2011) 353-361.
- [56] F.P. Barry, Mesenchymal stem cell therapy in joint disease, in: *Tissue Engineering of Cartilage and Bone: Novartis Foundation Symposium 249*, Wiley Online Library, 2003, pp. 86-102.
- [57] N.H. Goradel, F.G. Hour, B. Negahdari, Z.V. Malekshahi, M. Hashemzahi, A. Masoudifar, H.

- Mirzaei, Stem cell therapy: a new therapeutic option for cardiovascular diseases, *Journal of cellular biochemistry*, 119 (2018) 95-104.
- [58] F. Dazzi, M. Krampera, Mesenchymal stem cells and autoimmune diseases, *Best Practice & Research Clinical Haematology*, 24 (2011) 49-57.
- [59] J. Tolar, L. Xia, M.J. Riddle, C.J. Lees, C.R. Eide, R.T. McElmurry, M. Titeux, M.J. Osborn, T.C. Lund, A. Hovnanian, Induced pluripotent stem cells from individuals with recessive dystrophic epidermolysis bullosa, *Journal of Investigative Dermatology*, 131 (2011) 848-856.
- [60] N. Kim, S.-G. Cho, Clinical applications of mesenchymal stem cells, *The Korean journal of internal medicine*, 28 (2013) 387.
- [61] P.D. Rathjen, J. Lake, L. Whyatt, M.D. Bettess, J. Rathjen, Properties and uses of embryonic stem cells: prospects for application to human biology and gene therapy, *Reproduction, fertility and development*, 10 (1998) 31-48.
- [62] I.H. Schulman, V. Suncion, V. Karantalis, W. Balkan, J.M. Hare, C.C.T.R. Network, Clinical research skills development program in cell-based regenerative medicine, *Stem cells translational medicine*, 4 (2015) 118-122.
- [63] S. Liu, J. Zhou, X. Zhang, Y. Liu, J. Chen, B. Hu, J. Song, Y. Zhang, Strategies to optimize adult stem cell therapy for tissue regeneration, *International journal of molecular sciences*, 17 (2016) 982.
- [64] G. Shroff, J.K. Barthakur, Safety of human embryonic stem cells in patients with terminal/incurable conditions- a retrospective analysis, *Ann Neurosci*, 22 (2015) 132-138.
- [65] O.Y. Bang, E.H. Kim, J.M. Cha, G.J. Moon, Adult stem cell therapy for stroke: challenges and progress, *Journal of stroke*, 18 (2016) 256.
- [66] A.E. Omole, A.O.J. Fakoya, Ten years of progress and promise of induced pluripotent stem cells: historical origins, characteristics, mechanisms, limitations, and potential applications, *PeerJ*, 6 (2018) e4370.
- [67] K. Masuda, H. Kawamoto, Possible NK cell-mediated immune responses against iPSC-derived cells in allogeneic transplantation settings, *Inflammation and Regeneration*, 41 (2021) 1-9.
- [68] N.B. Vu, H.T. Nguyen, R. Palumbo, R. Pellicano, S. Fagoonee, P.V. Pham, Stem cell-derived exosomes for wound healing: current status and promising directions, *Minerva medica*, 112 (2020) 384-400.
- [69] C.A. Herberts, M.S. Kwa, H.P. Hermsen, Risk factors in the development of stem cell therapy, *Journal of translational medicine*, 9 (2011) 1-14.
- [70] M.D. Griffin, T. Ritter, B.P. Mahon, Immunological aspects of allogeneic mesenchymal stem cell therapies, *Human gene therapy*, 21 (2010) 1641-1655.
- [71] E. Rusu, L.G. Necula, A.I. Neagu, M. Alecu, C. Stan, R. Albulescu, C.P. Tanase, Current status of stem cell therapy: opportunities and limitations, *Turkish Journal of Biology*, 40 (2016) 955-967.
- [72] C. de Rham, J. Villard, Potential and limitation of HLA-based banking of human pluripotent stem cells for cell therapy, *Journal of immunology research*, 2014 (2014).
- [73] C. Li, S. Chen, Y. Zhou, Y. Zhao, P. Liu, J. Cai, Application of induced pluripotent stem cell transplants: Autologous or allogeneic?, *Life sciences*, 212 (2018) 145-149.
- [74] A. Kawamura, S. Miyagawa, S. Fukushima, T. Kawamura, N. Kashiya, E. Ito, T. Watabe, S. Masuda, K. Toda, J. Hatazawa, Teratocarcinomas arising from allogeneic induced pluripotent stem cell-derived cardiac tissue constructs provoked host immune rejection in mice, *Scientific reports*, 6 (2016) 19464.
- [75] M. Madrid, C. Sumen, S. Aivio, N. Saklayan, Autologous induced pluripotent stem cell-based cell

- therapies: promise, progress, and challenges, *Current protocols*, 1 (2021) e88.
- [76] Y. Teramura, H. Iwata, Bioartificial pancreas: Microencapsulation and conformal coating of islet of Langerhans, *Advanced drug delivery reviews*, 62 (2010) 827-840.
- [77] J. Iyer, P. Khayambashi, S.D. Tran, Salivary gland regeneration and repair in Sjögren's syndrome, in: *Translational Autoimmunity*, Elsevier, 2023, pp. 509-529.
- [78] D. Steffens, D.I. Braghirolli, N. Maurmann, P. Pranke, Update on the main use of biomaterials and techniques associated with tissue engineering, *Drug discovery today*, 23 (2018) 1474-1488.
- [79] U. Kneser, D.J. Schaefer, E. Polykandriotis, R.E. Horch, Tissue engineering of bone: the reconstructive surgeon's point of view, *Journal of cellular and molecular medicine*, 10 (2006) 7-19.
- [80] R. Santini, M.C. Vinci, S. Pandolfi, J.Y. Penachioni, V. Montagnani, B. Olivito, R. Gattai, N. Pimpinelli, G. Gerlini, L. Borgognoni, Hedgehog-Gli signaling drives self-renewal and tumorigenicity of human melanoma-initiating cells, *Stem cells*, 30 (2012) 1808-1818.
- [81] X. Zeng, C. Liu, J. Yao, H. Wan, G. Wan, Y. Li, N. Chen, Breast cancer stem cells, heterogeneity, targeting therapies and therapeutic implications, *Pharmacological research*, 163 (2021) 105320.
- [82] S. Naz, F.R. Khan, I. Khan, R.R. Zohra, A. Salim, N. Mohammed, T. Ahmad, Comparative analysis of dental pulp stem cells and stem cells from human exfoliated teeth in terms of growth kinetics, immunophenotype, self-renewal and multi lineage differentiation potential for future perspective of calcified tissue regeneration, *Pakistan Journal of Medical Sciences*, 38 (2022) 1228.
- [83] H. Lin, B. Wang, J. Yu, J. Wang, Q. Li, B. Cao, Protein arginine methyltransferase 8 gene enhances the colon cancer stem cell (CSC) function by upregulating the pluripotency transcription factor, *Journal of Cancer*, 9 (2018) 1394.
- [84] J. Tu, G. Tian, H.-H. Cheung, W. Wei, T.-I. Lee, Gas5 is an essential lncRNA regulator for self-renewal and pluripotency of mouse embryonic stem cells and induced pluripotent stem cells, *Stem cell research & therapy*, 9 (2018) 1-13.
- [85] B.C.M. Fernandes, P.D.M. Paes, C.G. Pereira, G. Silveira, Ten years of iPSC: clinical potential and advances in vitro hematopoietic differentiation.
- [86] Y. Yan, P. Yin, H. Gong, Y. Xue, G. Zhang, B. Fang, Z. Chen, Y. Li, C. Yang, Z. Huang, Nucleosome assembly protein 1-like 1 (Nap111) regulates the proliferation of murine induced pluripotent stem cells, *Cellular Physiology and Biochemistry*, 38 (2016) 340-350.
- [87] A. Vats, N. Tolley, A. Bishop, J. Polak, Embryonic stem cells and tissue engineering: delivering stem cells to the clinic, *Journal of the Royal Society of Medicine*, 98 (2005) 346-350.
- [88] H. Zhu, T. Kimura, S. Swami, J.Y. Wu, Pluripotent stem cells as a source of osteoblasts for bone tissue regeneration, *Biomaterials*, 196 (2019) 31-45.
- [89] S.C. Li, Y. Jin, W.G. Loudon, Y. Song, Z. Ma, L.P. Weiner, J.F. Zhong, Increase developmental plasticity of human keratinocytes with gene suppression, *Proceedings of the National Academy of Sciences*, 108 (2011) 12793-12798.
- [90] L. Gong, Q. Yan, Y. Zhang, X. Fang, B. Liu, X. Guan, Cancer cell reprogramming: a promising therapy converting malignancy to benignity, *Cancer Communications*, 39 (2019) 1-13.
- [91] C.-C. Ku, K. Wuputra, J.-B. Pan, C.-P. Li, C.-J. Liu, Y.-C. Liu, S. Saito, T.-F. Chan, C.-S. Lin, D.-C. Wu, Generation of human stomach cancer iPSC-derived organoids induced by *Helicobacter pylori* infection and their application to gastric cancer research, *Cells*, 11 (2022) 184.
- [92] H. Suh, Tissue restoration, tissue engineering and regenerative medicine, *Yonsei Med J*, 41 (2000) 681-684.

- [93] Y. Su, M.S. Toftdal, A. Le Friec, M. Dong, X. Han, M. Chen, 3D electrospun synthetic extracellular matrix for tissue regeneration, *Small Science*, 1 (2021) 2100003.
- [94] Z. Fereshteh, Freeze-drying technologies for 3D scaffold engineering, in: *Functional 3D tissue engineering scaffolds*, Elsevier, 2018, pp. 151-174.
- [95] B.L. Rodriguez, L.M. Larkin, Functional three-dimensional scaffolds for skeletal muscle tissue engineering, in: *Functional 3D Tissue Engineering Scaffolds*, Elsevier, 2018, pp. 279-304.
- [96] X. Hu, W. Zhang, X. Li, D. Zhong, Y. Li, J. Li, R. Jin, Strategies to modulate the redifferentiation of chondrocytes, *Frontiers in Bioengineering and Biotechnology*, 9 (2021) 764193.
- [97] Y. Lv, H. Wang, G. Li, B. Zhao, Three-dimensional decellularized tumor extracellular matrices with different stiffness as bioengineered tumor scaffolds, *Bioactive materials*, 6 (2021) 2767-2782.
- [98] U. Kneser, D. Schaefer, B. Munder, C. Klemm, C. Andree, G. Stark, Tissue engineering of bone, *Minimally Invasive Therapy & Allied Technologies*, 11 (2002) 107-116.
- [99] J.P. Vacanti, R. Langer, Tissue engineering: the design and fabrication of living replacement devices for surgical reconstruction and transplantation, *The lancet*, 354 (1999) S32-S34.
- [100] P.R. Baraniak, T.C. McDevitt, Stem cell paracrine actions and tissue regeneration, *Regenerative medicine*, 5 (2010) 121-143.
- [101] A. Bergmann, H. Steller, Apoptosis, stem cells, and tissue regeneration, *Science signaling*, 3 (2010) re8-re8.
- [102] Y. Tabata, Biomaterial technology for tissue engineering applications, *Journal of the Royal Society interface*, 6 (2009) S311-S324.
- [103] N.T. Feric, M. Radisic, Strategies and challenges to myocardial replacement therapy, *Stem cells translational medicine*, 5 (2016) 410-416.
- [104] G.I. Lambrou, E. Remboutsika, Proliferation versus regeneration: the good, the bad and the ugly, in: *Frontiers Media SA*, 2014, pp. 10.
- [105] A.V. Molofsky, R. Pardal, S.J. Morrison, Diverse mechanisms regulate stem cell self-renewal, *Current opinion in cell biology*, 16 (2004) 700-707.
- [106] T. Stiehl, A. Marciniak-Czochra, Stem cell self-renewal in regeneration and cancer: insights from mathematical modeling, *Current Opinion in Systems Biology*, 5 (2017) 112-120.
- [107] L.I. Zon, Intrinsic and extrinsic control of haematopoietic stem-cell self-renewal, *Nature*, 453 (2008) 306-313.
- [108] E. Fuchs, T. Chen, A matter of life and death: self - renewal in stem cells, *EMBO reports*, 14 (2013) 39-48.
- [109] L. Li, T. Xie, Stem cell niche: structure and function, *Annu. Rev. Cell Dev. Biol.*, 21 (2005) 605-631.
- [110] D.L. Jones, A.J. Wagers, No place like home: anatomy and function of the stem cell niche, *Nature reviews Molecular cell biology*, 9 (2008) 11-21.
- [111] S. Matsuda, T. Yan, A. Mizutani, T. Sota, Y. Hiramoto, M. Prieto - Vila, L. Chen, A. Satoh, T. Kudoh, T. Kasai, Cancer stem cells maintain a hierarchy of differentiation by creating their niche, *International journal of cancer*, 135 (2014) 27-36.
- [112] K.A. Becker, P.N. Ghule, J.A. Therrien, J.B. Lian, J.L. Stein, A.J. Van Wijnen, G.S. Stein, Self - renewal of human embryonic stem cells is supported by a shortened G1 cell cycle phase, *Journal of cellular physiology*, 209 (2006) 883-893.
- [113] J. Wray, T. Kalkan, A.G. Smith, The ground state of pluripotency, *Biochemical Society Transactions*, 38 (2010) 1027-1032.

- [114] M. Kinoshita, A. Smith, Pluripotency deconstructed, Development, growth & differentiation, 60 (2018) 44-52.
- [115] S.O. Min, S.W. Lee, S.Y. Bak, K.S. Kim, Ideal sphere-forming culture conditions to maintain pluripotency in a hepatocellular carcinoma cell lines, Cancer cell international, 15 (2015) 1-9.
- [116] P.S. Thiagarajan, M. Sinyuk, S.M. Turaga, E.E. Mulkearns-Hubert, J.S. Hale, V. Rao, A. Demelash, C. Saygin, A. China, T.J. Alban, Cx26 drives self-renewal in triple-negative breast cancer via interaction with NANOG and focal adhesion kinase, Nature communications, 9 (2018) 578.
- [117] D. Pedregal-Mallo, F. Hermida-Prado, R. Granda-Díaz, I. Montoro-Jiménez, E. Allonca, E. Pozo-Agundo, M. Álvarez-Fernández, C. Álvarez-Marcos, J.M. García-Pedrero, J.P. Rodrigo, Prognostic significance of the pluripotency factors NANOG, SOX2, and OCT4 in head and neck squamous cell carcinomas, Cancers, 12 (2020) 1794.
- [118] S. Yamanaka, Pluripotent stem cell-based cell therapy—promise and challenges, Cell stem cell, 27 (2020) 523-531.
- [119] E.M. Abdelalim, A. Bonnefond, A. Bennaceur-Griscelli, P. Froguel, Pluripotent stem cells as a potential tool for disease modelling and cell therapy in diabetes, Stem Cell Reviews and Reports, 10 (2014) 327-337.
- [120] S. Yamanaka, J. Li, G. Kania, S. Elliott, R.P. Wersto, J. Van Eyk, A.M. Wobus, K.R. Boheler, Pluripotency of embryonic stem cells, Cell and tissue research, 331 (2008) 5-22.
- [121] K.R. Boheler, Stem cell pluripotency: a cellular trait that depends on transcription factors, chromatin state and a checkpoint deficient cell cycle, Journal of cellular physiology, 221 (2009) 10-17.
- [122] A.E. Bishop, L.D. Buttery, J.M. Polak, Embryonic stem cells, The Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland, 197 (2002) 424-429.
- [123] S. Raab, M. Klingenstein, S. Liebau, L. Linta, A comparative view on human somatic cell sources for iPSC generation, Stem cells international, 2014 (2014).
- [124] S. Sun, R.R. White, K.E. Fischer, Z. Zhang, S.N. Austad, J. Vijg, Inducible aging in *Hydra oligactis* implicates sexual reproduction, loss of stem cells, and genome maintenance as major pathways, GeroScience, 42 (2020) 1119-1132.
- [125] S. Yamanaka, A fresh look at iPS cells, cell, 137 (2009) 13-17.
- [126] T. Halevy, A. Urbach, Comparing ESC and iPSC—based models for human genetic disorders, Journal of clinical medicine, 3 (2014) 1146-1162.
- [127] J.M. Iglesias, J. Gumuzio, A.G. Martin, Linking pluripotency reprogramming and cancer, Stem cells translational medicine, 6 (2017) 335-339.
- [128] T. Huang, X. Song, D. Xu, D. Tiek, A. Goenka, B. Wu, N. Sastry, B. Hu, S.-Y. Cheng, Stem cell programs in cancer initiation, progression, and therapy resistance, Theranostics, 10 (2020) 8721.
- [129] Y. Zhou, L. Xia, H. Wang, L. Oyang, M. Su, Q. Liu, J. Lin, S. Tan, Y. Tian, Q. Liao, Cancer stem cells in progression of colorectal cancer, Oncotarget, 9 (2018) 33403.
- [130] Y. Yang, J.-J. Yang, H. Tao, W.-S. Jin, New perspectives on β -catenin control of cell fate and proliferation in colon cancer, Food and chemical toxicology, 74 (2014) 14-19.
- [131] T. Mishina, H. Dosaka-Akita, I. Kinoshita, F. Hommura, T. Morikawa, H. Katoh, Y. Kawakami, Cyclin D1 expression in non-small-cell lung cancers: its association with altered p53 expression, cell proliferation and clinical outcome, British journal of cancer, 80 (1999) 1289-1295.
- [132] N. Rivlin, G. Koifman, V. Rotter, p53 orchestrates between normal differentiation and cancer, in: Seminars in cancer biology, Elsevier, 2015, pp. 10-17.

- [133] P.M. Evans, C. Liu, Roles of Krüppel - like factor 4 in normal homeostasis, cancer and stem cells, *Acta biochimica et biophysica Sinica*, 40 (2008) 554-564.
- [134] S. Yamanaka, K. Takahashi, Induction of pluripotent stem cells from mouse fibroblast cultures, *Tanpakushitsu kakusan koso. Protein, nucleic acid, enzyme*, 51 (2006) 2346-2351.
- [135] L. Yu, H. Wen, C. Liu, C. Wang, H. Yu, K. Zhang, Q. Han, Y. Liu, Z. Han, Z. Li, Embryonic stem cell-derived extracellular vesicles rejuvenate senescent cells and antagonize aging in mice, *Bioactive Materials*, 29 (2023) 85-97.
- [136] L.M. Eisenberg, C.A. Eisenberg, Stem cell plasticity, cell fusion, and transdifferentiation, *Birth Defects Research Part C: Embryo Today: Reviews*, 69 (2003) 209-218.
- [137] M.A. Nieto, Epithelial plasticity: a common theme in embryonic and cancer cells, *Science*, 342 (2013) 1234850.
- [138] G. Le Minh, M.J. Reginato, Role of O-GlcNAcylation on cancer stem cells: Connecting nutrient sensing to cell plasticity, *Advances in Cancer Research*, 157 (2023) 195-228.
- [139] Z.-D. Shi, K. Pang, Z.-X. Wu, Y. Dong, L. Hao, J.-X. Qin, W. Wang, Z.-S. Chen, C.-H. Han, Tumor cell plasticity in targeted therapy-induced resistance: mechanisms and new strategies, *Signal Transduction and Targeted Therapy*, 8 (2023) 113.
- [140] N.M. Warriar, N. Kelkar, C.T. Johnson, T. Govindarajan, V. Prabhu, P. Kumar, Understanding cancer stem cells and plasticity: Towards better therapeutics, *European Journal of Cell Biology*, (2023) 151321.
- [141] S. Temple, Stem cell plasticity—building the brain of our dreams, *Nature Reviews Neuroscience*, 2 (2001) 513-520.
- [142] S. Romanazzo, K. Lin, P. Srivastava, K.A. Kilian, Targeting cell plasticity for regeneration: From in vitro to in vivo reprogramming, *Adv Drug Deliv Rev*, 161-162 (2020) 124-144.
- [143] A. Das, M. Sinha, S. Datta, M. Abas, S. Chaffee, C.K. Sen, S. Roy, Monocyte and macrophage plasticity in tissue repair and regeneration, *Am J Pathol*, 185 (2015) 2596-2606.
- [144] M.R. Doherty, J.M. Smigiel, D.J. Junk, M.W. Jackson, Cancer stem cell plasticity drives therapeutic resistance, *Cancers*, 8 (2016) 8.
- [145] Y. Zhang, R.A. Weinberg, Epithelial-to-mesenchymal transition in cancer: complexity and opportunities, *Frontiers of medicine*, 12 (2018) 361-373.
- [146] F.S. Ramos, L. Wons, I.J. Cavalli, E. Ribeiro, Epithelial-mesenchymal transition in cancer: An overview, *Integr. Cancer Sci. Ther*, 4 (2017).
- [147] J.G. Gill, E.M. Langer, R.C. Lindsley, M. Cai, T.L. Murphy, M. Kyba, K.M. Murphy, Snail and the microRNA-200 family act in opposition to regulate epithelial-to-mesenchymal transition and germ layer fate restriction in differentiating ESCs, *Stem cells*, 29 (2011) 764-776.
- [148] M.A. Esteban, X. Bao, Q. Zhuang, T. Zhou, B. Qin, D. Pei, The mesenchymal-to-epithelial transition in somatic cell reprogramming, *Curr Opin Genet Dev*, 22 (2012) 423-428.
- [149] M. Takaishi, M. Tarutani, J. Takeda, S. Sano, Mesenchymal to Epithelial Transition Induced by Reprogramming Factors Attenuates the Malignancy of Cancer Cells, *PLoS One*, 11 (2016) e0156904.
- [150] G. Giannelli, P. Koudelkova, F. Dituri, W. Mikulits, Role of epithelial to mesenchymal transition in hepatocellular carcinoma, *J Hepatol*, 65 (2016) 798-808.
- [151] S. Singh, S. Brabletz, P. Arnold, M. Schicht, F. Paulsen, Epithelial-mesenchymal transition in the lacrimal gland morphogenesis, damage and repair, *Ocul Surf*, 29 (2023) 401-405.

- [152] P. Smeriglio, A. Zalc, Cranial Neural Crest Cells Contribution to Craniofacial Bone Development and Regeneration, *Curr Osteoporos Rep*, (2023) 1-8.
- [153] M.Z. Ratajczak, K. Bujko, A. Mack, M. Kucia, J. Ratajczak, Cancer from the perspective of stem cells and misappropriated tissue regeneration mechanisms, *Leukemia*, 32 (2018) 2519-2526.
- [154] Y. Wang, E. Bella, C.S. Lee, C. Migliaresi, L. Pelcastre, Z. Schwartz, B.D. Boyan, A. Motta, The synergistic effects of 3-D porous silk fibroin matrix scaffold properties and hydrodynamic environment in cartilage tissue regeneration, *Biomaterials*, 31 (2010) 4672-4681.
- [155] Y. Tabata, Tissue regeneration based on growth factor release, *Tissue engineering*, 9 (2003) 5-15.
- [156] L. Polo-Corrales, M. Latorre-Esteves, J.E. Ramirez-Vick, Scaffold design for bone regeneration, *Journal of nanoscience and nanotechnology*, 14 (2014) 15-56.
- [157] G.-M.L.P.M. Tian, L. Shamirzaei-Jeshvaghani E. Dehghani L. Ramakrsihna S. World J, *Stem Cells*, 7 (2015) 728.
- [158] P. Koria, Delivery of growth factors for tissue regeneration and wound healing, *BioDrugs*, 26 (2012) 163-175.
- [159] C. Frantz, K.M. Stewart, V.M. Weaver, The extracellular matrix at a glance, *Journal of cell science*, 123 (2010) 4195-4200.
- [160] I.L.S. Chua, H.-W. Kim, J.H. Lee, Signaling of extracellular matrices for tissue regeneration and therapeutics, *Tissue engineering and regenerative medicine*, 13 (2016) 1-12.
- [161] K.S. Midwood, L.V. Williams, J.E. Schwarzbauer, Tissue repair and the dynamics of the extracellular matrix, *The international journal of biochemistry & cell biology*, 36 (2004) 1031-1037.
- [162] W.P. Daley, S.B. Peters, M. Larsen, Extracellular matrix dynamics in development and regenerative medicine, *Journal of cell science*, 121 (2008) 255-264.
- [163] S. Yi, F. Ding, L. Gong, X. Gu, Extracellular matrix scaffolds for tissue engineering and regenerative medicine, *Current stem cell research & therapy*, 12 (2017) 233-246.
- [164] E.L. Fong, S.E. Lamhamedi-Cherradi, E. Burdett, V. Ramamoorthy, A.J. Lazar, F.K. Kasper, M.C. Farach-Carson, D. Vishwamitra, E.G. Demicco, B.A. Menegaz, H.M. Amin, A.G. Mikos, J.A. Ludwig, Modeling Ewing sarcoma tumors in vitro with 3D scaffolds, *Proc Natl Acad Sci U S A*, 110 (2013) 6500-6505.
- [165] C.Y. Liaw, S. Ji, M. Guvendiren, Engineering 3D Hydrogels for Personalized In Vitro Human Tissue Models, *Adv Healthc Mater*, 7 (2018) 1701165.
- [166] H. Cui, W. Zhu, B. Holmes, L.G. Zhang, Biologically Inspired Smart Release System Based on 3D Bioprinted Perfused Scaffold for Vascularized Tissue Regeneration, *Adv Sci (Weinh)*, 3 (2016) 1600058.
- [167] O. Buhome, M. Wongwattanakul, J. Daduang, T. Limpai boon, 3D Silk Fibroin-Gelatin/Hyaluronic Acid/Heparan sulfate scaffold enhances expression of stemness and EMT markers in Cholangiocarcinoma, in vivo, 36 (2022) 1155-1167.
- [168] M. Paolillo, R. Colombo, M. Serra, L. Belvisi, A. Papetti, E. Ciusani, S. Comincini, S. Schinelli, Stem-like cancer cells in a dynamic 3D culture system: a model to study metastatic cell adhesion and anti-cancer drugs, *Cells*, 8 (2019) 1434.
- [169] M.V. Giolito, L. Claret, C. Frau, M. Plateroti, A Three-dimensional model of spheroids to study colon cancer stem cells, *JoVE (Journal of Visualized Experiments)*, (2021) e61783.
- [170] M. Houshmand, M. Soleimani, A. Atashi, G. Saglio, M. Abdollahi, M. Nikougoftar Zarif, Mimicking the acute myeloid leukemia niche for

- molecular study and drug screening, *Tissue Engineering Part C: Methods*, 23 (2017) 72-85.
- [171] F.M. Kievit, S.J. Florczyk, M.C. Leung, K. Wang, J.D. Wu, J.R. Silber, R.G. Ellenbogen, J.S. Lee, M. Zhang, Proliferation and enrichment of CD133+ glioblastoma cancer stem cells on 3D chitosan-alginate scaffolds, *Biomaterials*, 35 (2014) 9137-9143.
- [172] M. Leung, F.M. Kievit, S.J. Florczyk, O. Veiseh, J. Wu, J.O. Park, M. Zhang, Chitosan-alginate scaffold culture system for hepatocellular carcinoma increases malignancy and drug resistance, *Pharmaceutical research*, 27 (2010) 1939-1948.
- [173] J. Lopez, M. Ruiz-Toranzo, C. Antich, C. Chocarro-Wrona, E. López-Ruiz, G. Jiménez, J. Marchal, Biofabrication of a Tri-layered 3D-Bioprinted CSC-based Malignant Melanoma Model for Personalized Cancer Treatment, *Biofabrication*, (2022).
- [174] S. Feng, X. Duan, P.-K. Lo, S. Liu, X. Liu, H. Chen, Q. Wang, Expansion of breast cancer stem cells with fibrous scaffolds, *Integrative Biology*, 5 (2013) 768-777.
- [175] K.A. Fitzgerald, J. Guo, E.G. Tierney, C.M. Curtin, M. Malhotra, R. Darcy, F.J. O'Brien, C.M. O'Driscoll, The use of collagen-based scaffolds to simulate prostate cancer bone metastases with potential for evaluating delivery of nanoparticulate gene therapeutics, *Biomaterials*, 66 (2015) 53-66.
- [176] M. Bordoni, E. Karabulut, V. Kuzmenko, V. Fantini, O. Pansarasa, C. Cereda, P. Gatenholm, 3D printed conductive nanocellulose scaffolds for the differentiation of human neuroblastoma cells, *Cells*, 9 (2020) 682.
- [177] S. Braccini, C. Tacchini, F. Chiellini, D. Puppi, Polymeric hydrogels for in vitro 3D ovarian cancer modeling, *International Journal of Molecular Sciences*, 23 (2022) 3265.
- [178] E. Tottoli, R. Dorati, I. Genta, E. Chiesa, S. Pisani, B. Conti, Skin wound healing process and new emerging technologies for skin wound care and regeneration. *Pharmaceutics*. 2020; 12 (8): 735, in.
- [179] L. Suamte, A. Tirkey, J. Barman, P.J. Babu, Various manufacturing methods and ideal properties of scaffolds for tissue engineering applications, *Smart Materials in Manufacturing*, 1 (2023) 100011.
- [180] M. Heinrich, A. Mostafa, J. Morton, L. Hawinkels, J. Prakash, Translating Complexity and Heterogeneity of Pancreatic Tumor: 3D, in, *Vitro*, 2021.
- [181] O.A. Sandiford, R.J. Donnelly, M.H. El-Far, L.M. Burgmeyer, G. Sinha, S.H. Pamarthi, L.S. Sherman, A.I. Ferrer, D.E. DeVore, S.A. Patel, Mesenchymal stem cell-secreted extracellular vesicles instruct stepwise dedifferentiation of breast cancer cells into dormancy at the bone marrow perivascular region, *Cancer research*, 81 (2021) 1567-1582.
- [182] M.F. Arisi, R.A. Starker, S. Addya, Y. Huang, S.V. Fernandez, All trans-retinoic acid (ATRA) induces re-differentiation of early transformed breast epithelial cells, *International journal of oncology*, 44 (2014) 1831-1842.
- [183] M. Yan, Q. Liu, Differentiation therapy: a promising strategy for cancer treatment, *Chinese journal of cancer*, 35 (2016) 1-3.
- [184] I.N. Shvemberger, Conversion of malignant cells into normal ones, *International review of cytology*, 103 (1986) 341-386.
- [185] J.W. Bachman, T. Hillen, Mathematical optimization of the combination of radiation and differentiation therapies for cancer, *Frontiers in oncology*, 3 (2013) 42398.
- [186] G.B. Pierce, C. Wallace, Differentiation of malignant to benign cells, *Cancer Res*, 31 (1971) 127-134.
- [187] E. Mueller, P. Sarraf, P. Tontonoz, R.M. Evans, K.J. Martin, M. Zhang, C. Fletcher, S. Singer, B.M. Spiegelman, Terminal differentiation of human

- breast cancer through PPAR γ , *Molecular cell*, 1 (1998) 465-470.
- [188] M. Huang, C. Chen, J. Geng, D. Han, T. Wang, T. Xie, L. Wang, Y. Wang, C. Wang, Z. Lei, Targeting KDM1A attenuates Wnt/ β -catenin signaling pathway to eliminate sorafenib-resistant stem-like cells in hepatocellular carcinoma, *Cancer letters*, 398 (2017) 12-21.
- [189] H. Kawamata, M. Tachibana, T. Fujimori, Y. Imai, Differentiation-inducing therapy for solid tumors, *Current pharmaceutical design*, 12 (2006) 379-385.
- [190] B.X. Hoang, B. Han, W.H. Fang, H.D. Tran, C. Hoang, D.G. Shaw, T.Q. Nguyen, The rationality of implementation of dimethyl sulfoxide as differentiation-inducing agent in cancer therapy, *Cancer Diagnosis & Prognosis*, 3 (2023) 1.
- [191] M.T. Shekhani, A.S. Jayanthi, N. Maddodi, V. Setaluri, Cancer stem cells and tumor transdifferentiation: implications for novel therapeutic strategies, *Am J Stem Cells*, 2 (2013) 52-61.
- [192] D. Fang, T.K. Nguyen, K. Leishear, R. Finko, A.N. Kulp, S. Hotz, P.A. Van Belle, X. Xu, D.E. Elder, M. Herlyn, A tumorigenic subpopulation with stem cell properties in melanomas, *Cancer research*, 65 (2005) 9328-9337.
- [193] J. Song, H. Zhou, D. Gu, Y. Xu, Hepatocellular carcinoma differentiation: Research progress in mechanism and treatment, *Frontiers in Oncology*, 11 (2022) 790358.
- [194] M. Tatematsu, Y. Nagamine, E. Farber, Redifferentiation as a basis for remodeling of carcinogen-induced hepatocyte nodules to normal appearing liver, *Cancer research*, 43 (1983) 5049-5058.
- [195] F. Pezzini, L. Bettinetti, F. Di Leva, M. Bianchi, E. Zoratti, R. Carrozzo, F.M. Santorelli, M. Delledonne, M. Lalowski, A. Simonati, Transcriptomic profiling discloses molecular and cellular events related to neuronal differentiation in SH-SY5Y neuroblastoma cells, *Cellular and molecular neurobiology*, 37 (2017) 665-682.
- [196] J.R. Murillo, L. Goto-Silva, A. Sánchez, F.C. Nogueira, G.B. Domont, M. Junqueira, Quantitative proteomic analysis identifies proteins and pathways related to neuronal development in differentiated SH-SY5Y neuroblastoma cells, *EuPA open proteomics*, 16 (2017) 1-11.
- [197] E. Abemayor, N. Sidell, Human neuroblastoma cell lines as models for the in vitro study of neoplastic and neuronal cell differentiation, *Environmental health perspectives*, 80 (1989) 3-15.
- [198] S. Azzi, S. Bruno, J. Giron-Michel, D. Clay, A. Devocelle, M. Croce, S. Ferrini, S. Chouaib, A. Vazquez, B. Charpentier, Differentiation therapy: targeting human renal cancer stem cells with interleukin 15, *Journal of the National Cancer Institute*, 103 (2011) 1884-1898.
- [199] Y. Arima, H. Nobusue, H. Saya, Targeting of cancer stem cells by differentiation therapy, *Cancer science*, 111 (2020) 2689-2695.
- [200] Z. YangQJ, All-transretinoic acid inhibitstumorgrowthofhumanosteosarcoma, byactivating Smad signaling-inducedosteogenicdifferentiation, *IntJ Oncol*, 41 (2012) 153-160.
- [201] Z. Cheng, Z. He, Y. Cai, C. Zhang, G. Fu, H. Li, W. Sun, C. Liu, X. Cui, B. Ning, Conversion of hepatoma cells to hepatocyte-like cells by defined hepatocyte nuclear factors, *Cell research*, 29 (2019) 124-135.
- [202] D. Ishay-Ronen, M. Diepenbruck, R.K.R. Kalathur, N. Sugiyama, S. Tiede, R. Ivanek, G. Bantug, M.F. Morini, J. Wang, C. Hess, Gain fat—lose metastasis: converting invasive breast cancer cells into adipocytes inhibits cancer metastasis, *Cancer cell*, 35 (2019) 17-32. e16.
- [203] S. Roszkowski, Application of Polyphenols and Flavonoids in Oncological Therapy, *Molecules*, 28 (2023) 4080.

- [204] D. Avtanski, L. Poretsky, Phyto-polyphenols as potential inhibitors of breast cancer metastasis, *Molecular Medicine*, 24 (2018) 1-17.
- [205] E.S. Mocarski, J.W. Upton, W.J. Kaiser, Viral infection and the evolution of caspase 8-regulated apoptotic and necrotic death pathways, *Nature Reviews Immunology*, 12 (2012) 79-88.
- [206] M. Pei, J. Seidel, G. Vunjak-Novakovic, L. Freed, Growth factors for sequential cellular de-and re-differentiation in tissue engineering, *Biochemical and Biophysical Research Communications*, 294 (2002) 149-154.
- [207] C. Massard, E. Deutsch, J.-C. Soria, Tumour stem cell-targeted treatment: elimination or differentiation, *Annals of oncology*, 17 (2006) 1620-1624.
- [208] N. Ichiryu, P.J. Fairchild, Immune privilege of stem cells, *Embryonic Stem Cell Immunobiology: Methods and Protocols*, (2013) 1-16.
- [209] K. Mortezaee, Immune escape: A critical hallmark in solid tumors, *Life sciences*, 258 (2020) 118110.
- [210] B. Seliger, C. Massa, B. Yang, D. Bethmann, M. Kappler, A.W. Eckert, C. Wickenhauser, Immune escape mechanisms and their clinical relevance in head and neck squamous cell carcinoma, *International journal of molecular sciences*, 21 (2020) 7032.
- [211] I. Galea, I. Bechmann, V.H. Perry, What is immune privilege (not)?, *Trends in immunology*, 28 (2007) 12-18.
- [212] T.J. Miller, M.J. McCoy, C. Hemmings, B. Iacopetta, C.F. Platell, Expression of PD-L1 and SOX2 during rectal tumourigenesis: Potential mechanisms for immune escape and tumour cell invasion, *Oncology letters*, 16 (2018) 5761-5768.
- [213] K. Staveley-O'Carroll, E. Sotomayor, J. Montgomery, I. Borrello, L. Hwang, S. Fein, D. Pardoll, H. Levitsky, Induction of antigen-specific T cell anergy: an early event in the course of tumor progression, *Proceedings of the National Academy of Sciences*, 95 (1998) 1178-1183.
- [214] K.J. Wood, A. Bushell, J. Hester, Regulatory immune cells in transplantation, *Nature Reviews Immunology*, 12 (2012) 417-430.
- [215] M. Iero, R. Valenti, V. Huber, P. Filipazzi, G. Parmiani, S. Fais, L. Rivoltini, Tumour-released exosomes and their implications in cancer immunity, *Cell Death & Differentiation*, 15 (2008) 80-88.
- [216] F. Pierini, T.L. Lenz, Divergent allele advantage at human MHC genes: signatures of past and ongoing selection, *Molecular Biology and Evolution*, 35 (2018) 2145-2158.
- [217] J.F. Kaufman, C. Auffray, A.J. Korman, D.A. Shackelford, J. Strominger, The class II molecules of the human and murine major histocompatibility complex, *Cell*, 36 (1984) 1-13.
- [218] M.A. Ayala García, B. González Yebra, A.L. López Flores, E. Guaní Guerra, The major histocompatibility complex in transplantation, *Journal of transplantation*, 2012 (2012).
- [219] T. Yamaguchi, J.M. Dijkstra, Major histocompatibility complex (MHC) genes and disease resistance in fish, *Cells*, 8 (2019) 378.
- [220] C. Wedekind, D. Penn, MHC genes, body odours, and odour preferences, *Nephrology Dialysis Transplantation*, 15 (2000) 1269-1271.
- [221] J.G. Sambrook, F. Figueroa, S. Beck, A genome-wide survey of Major Histocompatibility Complex (MHC) genes and their paralogues in zebrafish, *Bmc Genomics*, 6 (2005) 1-10.
- [222] J. Trowsdale, J.C. Knight, Major histocompatibility complex genomics and human disease, *Annual review of genomics and human genetics*, 14 (2013) 301-323.
- [223] P. Marrack, J. Kappler, The T cell receptor, *Science*, 238 (1987) 1073-1079.

- [224] L.L. Lanier, Follow the leader: NK cell receptors for classical and nonclassical MHC class I, *Cell*, 92 (1998) 705-707.
- [225] C. Galassi, M. Musella, N. Manduca, E. Maccafeo, A. Sistigu, The immune privilege of cancer stem cells: a key to understanding tumor immune escape and therapy failure, *Cells*, 10 (2021) 2361.
- [226] H. Dianat-Moghadam, M. Sharifi, R. Salehi, M. Keshavarz, M. Shahgolzari, Z. Amoozgar, Engaging stemness improves cancer immunotherapy, *Cancer Letters*, (2022) 216007.
- [227] M.A. Nengroo, A. Verma, D. Datta, Cytokine chemokine network in tumor microenvironment: Impact on CSC properties and therapeutic applications, *Cytokine*, 156 (2022) 155916.
- [228] I.L. Sargent, A.M. Borzychowski, C.W. Redman, NK cells and human pregnancy—an inflammatory view, *Trends in immunology*, 27 (2006) 399-404.
- [229] P. Menendez, C. Bueno, L. Wang, M. Bhatia, Human embryonic stem cells: potential tool for achieving immunotolerance?, *Stem cell reviews*, 1 (2005) 151-158.
- [230] A. Wu, S. Wiesner, J. Xiao, K. Ericson, W. Chen, W.A. Hall, W.C. Low, J.R. Ohlfest, Expression of MHC I and NK ligands on human CD133+ glioma cells: possible targets of immunotherapy, *Journal of neuro-oncology*, 83 (2007) 121-131.
- [231] E. Shklovskaya, H. Rizos, MHC class I deficiency in solid tumors and therapeutic strategies to overcome it, *International journal of molecular sciences*, 22 (2021) 6741.
- [232] N. Badrinath, S.Y. Yoo, Recent advances in cancer stem cell-targeted immunotherapy, *Cancers*, 11 (2019) 310.
- [233] J. Bischof, M.-A. Westhoff, J.E. Wagner, M.-E. Halatsch, S. Trentmann, U. Knippschild, C.R. Wirtz, T. Burster, Cancer stem cells: The potential role of autophagy, proteolysis, and cathepsins in glioblastoma stem cells, *Tumor Biology*, 39 (2017) 1010428317692227.
- [234] A. Shokouhifar, J. Firouzi, M. Nouri, G.A. Sarab, M. Ebrahimi, NK cell upraise in the dark world of cancer stem cells, *Cancer Cell International*, 21 (2021) 1-15.
- [235] R. Apps, L. Gardner, A. Moffett, A critical look at HLA-G, *Trends in immunology*, 29 (2008) 313-321.
- [236] E.D. Carosella, P. Moreau, J. LeMaoult, N. Rouas-Freiss, HLA-G: from biology to clinical benefits, *Trends in immunology*, 29 (2008) 125-132.
- [237] N. Rouas-Freiss, P. Moreau, C. Menier, E.D. Carosella, HLA-G in cancer: a way to turn off the immune system, in: *Seminars in cancer biology*, Elsevier, 2003, pp. 325-336.
- [238] A. Lin, W.H. Yan, Intercellular transfer of HLA - G: Its potential in cancer immunology, *Clinical & Translational Immunology*, 8 (2019) e1077.
- [239] A. Desai, Y. Yan, S.L. Gerson, Concise reviews: cancer stem cell targeted therapies: toward clinical success, *Stem Cells Translational Medicine*, 8 (2019) 75-81.
- [240] F. Morandi, I. Airoidi, HLA-G and other immune checkpoint molecules as targets for novel combined immunotherapies, *International Journal of Molecular Sciences*, 23 (2022) 2925.
- [241] D. Geh, J. Leslie, R. Rumney, H.L. Reeves, T.G. Bird, D.A. Mann, Neutrophils as potential therapeutic targets in hepatocellular carcinoma, *Nature reviews Gastroenterology & hepatology*, 19 (2022) 257-273.
- [242] M. Zhou, C. Wang, S. Lu, Y. Xu, Z. Li, H. Jiang, Y. Ma, Tumor-associated macrophages in cholangiocarcinoma: complex interplay and potential therapeutic target, *EBioMedicine*, 67 (2021).

- [243] C. Li, P. Jiang, S. Wei, X. Xu, J. Wang, Regulatory T cells in tumor microenvironment: new mechanisms, potential therapeutic strategies and future prospects, *Molecular cancer*, 19 (2020) 1-23.
- [244] F. Veglia, A. Hashimoto, H. Dweep, E. Sanseviero, A. De Leo, E. Tcyganov, A. Kossenkov, C. Mulligan, B. Nam, G. Masters, Analysis of classical neutrophils and polymorphonuclear myeloid-derived suppressor cells in cancer patients and tumor-bearing mice, *Journal of Experimental Medicine*, 218 (2021).
- [245] T. Relation, M. Dominici, E.M. Horwitz, Concise review: an (im) penetrable shield: how the tumor microenvironment protects cancer stem cells, *Stem Cells*, 35 (2017) 1123-1130.
- [246] C. Sinha, L.C. Cunningham, An overview of the potential strategies for NK cell - based immunotherapy for acute myeloid leukemia, *Pediatric Blood & Cancer*, 63 (2016) 2078-2085.
- [247] S. Duan, W. Guo, Z. Xu, Y. He, C. Liang, Y. Mo, Y. Wang, F. Xiong, C. Guo, Y. Li, Natural killer group 2D receptor and its ligands in cancer immune escape, *Molecular cancer*, 18 (2019) 1-14.
- [248] B. Wang, Q. Wang, Z. Wang, J. Jiang, S.-C. Yu, Y.-F. Ping, J. Yang, S.-L. Xu, X.-Z. Ye, C. Xu, Metastatic consequences of immune escape from NK cell cytotoxicity by human breast cancer stem cells, *Cancer research*, 74 (2014) 5746-5757.
- [249] T. Bald, M.F. Krummel, M.J. Smyth, K.C. Barry, The NK cell–cancer cycle: advances and new challenges in NK cell–based immunotherapies, *Nature immunology*, 21 (2020) 835-847.
- [250] M. Tucci, S. Stucci, A. Passarelli, G. Giudice, F. Dammacco, F. Silvestris, The immune escape in melanoma: role of the impaired dendritic cell function, *Expert review of clinical immunology*, 10 (2014) 1395-1404.
- [251] P. Seeger, T. Musso, S. Sozzani, The TGF- β superfamily in dendritic cell biology, *Cytokine & Growth Factor Reviews*, 26 (2015) 647-657.
- [252] V. Triaca, V. Carito, E. Fico, P. Rosso, M. Fiore, M. Ralli, A. Lambiase, A. Greco, P. Tirassa, Cancer stem cells-driven tumor growth and immune escape: The Janus face of neurotrophins, *Aging (Albany NY)*, 11 (2019) 11770.
- [253] R.D. Stout, J. Suttles, Functional plasticity of macrophages: reversible adaptation to changing microenvironments, *Journal of leukocyte biology*, 76 (2004) 509-513.
- [254] G.S. Ashcroft, Bidirectional regulation of macrophage function by TGF- β , *Microbes and infection*, 1 (1999) 1275-1282.
- [255] C. Rébé, F. Végran, H. Berger, F. Ghiringhelli, STAT3 activation: A key factor in tumor immunoescape, *Jak-stat*, 2 (2013) e23010.
- [256] H. Lee, A.J. Jeong, S.-K. Ye, Highlighted STAT3 as a potential drug target for cancer therapy, *BMB reports*, 52 (2019) 415.
- [257] E. Tcyganov, J. Mastio, E. Chen, D.I. Gabrilovich, Plasticity of myeloid-derived suppressor cells in cancer, *Current opinion in immunology*, 51 (2018) 76-82.
- [258] K. Li, H. Shi, B. Zhang, X. Ou, Q. Ma, Y. Chen, P. Shu, D. Li, Y. Wang, Myeloid-derived suppressor cells as immunosuppressive regulators and therapeutic targets in cancer, *Signal Transduction and Targeted Therapy*, 6 (2021) 362.
- [259] M.A. Olivares-Urbano, C. Griñán-Lisón, J.A. Marchal, M.I. Núñez, CSC radioresistance: a therapeutic challenge to improve radiotherapy effectiveness in cancer, *Cells*, 9 (2020) 1651.
- [260] C. Tecchio, P. Scapini, G. Pizzolo, M.A. Cassatella, On the cytokines produced by human neutrophils in tumors, in: *Seminars in cancer biology*, Elsevier, 2013, pp. 159-170.
- [261] A.D. Gregory, A. McGarry Houghton, Tumor-associated neutrophils: new targets for cancer therapy, *Cancer research*, 71 (2011) 2411-2416.

- [262] A.M. Houghton, The paradox of tumor-associated neutrophils: fueling tumor growth with cytotoxic substances, *Cell cycle*, 9 (2010) 1732-1737.
- [263] L.W. Treffers, I.H. Hiemstra, T.W. Kuijpers, T.K. Van den Berg, H.L. Matlung, Neutrophils in cancer, *Immunological reviews*, 273 (2016) 312-328.
- [264] M.E. Shaul, Z.G. Fridlender, The dual role of neutrophils in cancer, in: *Seminars in Immunology*, Elsevier, 2021, pp. 101582.
- [265] H. Ikeda, K. Chamoto, T. Tsuji, Y. Suzuki, D. Wakita, T. Takeshima, T. Nishimura, The critical role of type - 1 innate and acquired immunity in tumor immunotherapy, *Cancer science*, 95 (2004) 697-703.
- [266] V.K.N. Hermann-Kleiter, Beyond CTLA-4 and PD-1: Orphan nuclear receptor, *Biochem. Biophys. Res. Commun*, 384 (2009) 405-408.
- [267] G. Giganti, M. Atif, Y. Mohseni, D. Mastronicola, N. Grageda, G.A. Povoleri, M. Miyara, C. Scottà, Treg cell therapy: How cell heterogeneity can make the difference, *European Journal of Immunology*, 51 (2021) 39-55.
- [268] A. O'Garra, P.L. Vieira, P. Vieira, A.E. Goldfeld, IL-10-producing and naturally occurring CD4+ Tregs: limiting collateral damage, *The Journal of clinical investigation*, 114 (2004) 1372-1378.
- [269] J. Li, X. Li, Q. Guo, Drug resistance in cancers: A free pass for bullying, *Cells*, 11 (2022) 3383.
- [270] Y. Xu, Y. Mao, Y. Lv, W. Tang, J. Xu, B cells in tumor metastasis: friend or foe?, *International Journal of Biological Sciences*, 19 (2023) 2382.
- [271] A. Kessel, T. Haj, R. Peri, A. Snir, D. Melamed, E. Sabo, E. Toubi, Human CD19+ CD25high B regulatory cells suppress proliferation of CD4+ T cells and enhance Foxp3 and CTLA-4 expression in T-regulatory cells, *Autoimmunity reviews*, 11 (2012) 670-677.
- [272] C.-Y. Huang, C.-L. Liu, C.-Y. Ting, Y.-T. Chiu, Y.-C. Cheng, M.W. Nicholson, P.C. Hsieh, Human iPSC banking: barriers and opportunities, *Journal of biomedical science*, 26 (2019) 1-14.
- [273] S. Solomon, F. Pitossi, M.S. Rao, Banking on iPSC-is it doable and is it worthwhile, *Stem Cell Reviews and Reports*, 11 (2015) 1-10.
- [274] E. Garreta, S. Sanchez, J. Lajara, N. Montserrat, J.C.I. Belmonte, Roadblocks in the Path of iPSC to the Clinic, *Current transplantation reports*, 5 (2018) 14-18.
- [275] J. Thornton, M. Edwards, W. Taylor, D. Barlow, Location of 'continuous' antigenic determinants in the protruding regions of proteins, *The EMBO journal*, 5 (1986) 409-413.
- [276] M. Sarmiento, P. Ramírez, R. Parody, M. Salas, N. Beffermann, V. Jara, P. Bertín, I. Pizarro, C. Lorca, E. Rivera, Advantages of non-cryopreserved autologous hematopoietic stem cell transplantation against a cryopreserved strategy, *Bone Marrow Transplantation*, 53 (2018) 960-966.
- [277] K. Takahashi, S. Yamanaka, Induced pluripotent stem cells in medicine and biology, *Development*, 140 (2013) 2457-2461.
- [278] M. Pakzad, S.N. Hassani, F. Abbasi, E. Hajizadeh-Saffar, L. Taghiyar, N. Fallah, N. Haghparast, A. Samadian, M. Ganjibakhsh, M. Dominici, A roadmap for the production of a GMP-compatible cell bank of allogeneic bone marrow-derived clonal mesenchymal stromal cells for cell therapy applications, *Stem Cell Reviews and Reports*, 18 (2022) 2279-2295.
- [279] E.W. Petersdorf, In celebration of Ruggero Ceppellini: HLA in transplantation, *Hla*, 89 (2017) 71-76.
- [280] E.M. Mickelson, E. Petersdorf, C. Anasetti, P. Martin, A. Woolfrey, J.A. Hansen, HLA matching in hematopoietic cell transplantation, *Human immunology*, 61 (2000) 92-100.
- [281] A. Shamsian, R. Sahebnaasagh, A. Norouzy, S.H. Hussein, M.H. Ghahremani, Z. Azizi, *Cancer*

- cells as a new source of induced pluripotent stem cells, *Stem Cell Research & Therapy*, 13 (2022) 459.
- [282] K. Izgi, H. Canatan, B. Iskender, Current status in cancer cell reprogramming and its clinical implications, *Journal of cancer research and clinical oncology*, 143 (2017) 371-383.
- [283] W. Fang, T. Zhou, H. Shi, M. Yao, D. Zhang, H. Qian, Q. Zeng, Y. Wang, F. Jin, C. Chai, Progranulin induces immune escape in breast cancer via up-regulating PD-L1 expression on tumor-associated macrophages (TAMs) and promoting CD8+ T cell exclusion, *Journal of Experimental & Clinical Cancer Research*, 40 (2021) 1-11.
- [284] E. Rouzbahani, J. Majidpoor, S. Najafi, K. Mortezaee, Cancer stem cells in immunoregulation and bypassing anti-checkpoint therapy, *Biomedicine & Pharmacotherapy*, 156 (2022) 113906.
- [285] D. Sarkar, I.V. Lebedeva, P. Gupta, L. Emdad, M. Sauane, P. Dent, D.T. Curiel, P.B. Fisher, Melanoma differentiation associated gene-7 (mda-7)/IL-24: a 'magic bullet' for cancer therapy?, *Expert opinion on biological therapy*, 7 (2007) 577-586.
- [286] H. Karvonen, M. Arjama, L. Kaleva, W. Niininen, H. Barker, R. Koivisto-Korander, J. Tapper, P. Pakarinen, H. Lassus, M. Loukovaara, Glucocorticoids induce differentiation and chemoresistance in ovarian cancer by promoting ROR1-mediated stemness, *Cell death & disease*, 11 (2020) 790.
- [287] C. Bailly, X. Thuru, B. Quesnel, Combined cytotoxic chemotherapy and immunotherapy of cancer: modern times, *NAR cancer*, 2 (2020) zcaa002.
- [288] K. Petrie, A. Zelent, S. Waxman, Differentiation therapy of acute myeloid leukemia: past, present and future, *Current opinion in hematology*, 16 (2009) 84-91.
- [289] D. Nowak, D. Stewart, H.P. Koefler, Differentiation therapy of leukemia: 3 decades of development, *Blood, The Journal of the American Society of Hematology*, 113 (2009) 3655-3665.
- [290] J. Jin, F. Grigore, C.C. Chen, M. Li, Self-renewal signaling pathways and differentiation therapies of glioblastoma stem cells, *International journal of oncology*, 59 (2021) 1-11.
- [291] K.H. Jung, J. Zhang, C. Zhou, H. Shen, M. Gagea, C. Rodriguez - Aguayo, G. Lopez - Berestein, A.K. Sood, L. Beretta, Differentiation therapy for hepatocellular carcinoma: Multifaceted effects of miR - 148a on tumor growth and phenotype and liver fibrosis, *Hepatology*, 63 (2016) 864-879.
- [292] A. Plotnikov, N. Kozler, G. Cohen, S. Carvalho, S. Duberstein, O. Almog, L.J. Solmesky, K.A. Shurrush, I. Babaev, S. Benjamin, PRMT1 inhibition induces differentiation of colon cancer cells, *Scientific Reports*, 10 (2020) 20030.
- [293] P.V. Pham, N.L. Phan, N.T. Nguyen, N.H. Truong, T.T. Duong, D.V. Le, K.D. Truong, N.K. Phan, Differentiation of breast cancer stem cells by knockdown of CD44: promising differentiation therapy, *Journal of translational medicine*, 9 (2011) 1-13.
- [294] G. Shao, W. Lai, X. Wan, J. Xue, Y. Wei, J. Jin, L. Zhang, Q. Lin, Q. Shao, S. Zou, Inactivation of EGFR/AKT signaling enhances TSA-induced ovarian cancer cell differentiation, *Oncology reports*, 37 (2017) 2891-2896.
- [295] E. Brambilla, D. Moro, S. Gazzeri, P. Brichon, H. Nagy-Mignotte, F. Morel, M. Jacrot, C. Brambilla, Cytotoxic chemotherapy induces cell differentiation in small-cell lung carcinoma, *Journal of Clinical Oncology*, 9 (1991) 50-61.
- [296] Z. Jin, Y. Lu, Y. Wu, J. Che, X. Dong, Development of differentiation modulators and targeted agents for treating neuroblastoma, *European Journal of Medicinal Chemistry*, 207 (2020) 112818.
- [297] P. Soballe, M. Herlyn, Cellular pathways leading to melanoma differentiation: therapeutic implications, *Melanoma Research*, 4 (1994) 213-223.

- [298] Y. Hagiwara, T. Kasukabe, Y. Kaneko, N. Niitsu, J. Okabe-Kado, Ellagic acid, a natural polyphenolic compound, induces apoptosis and potentiates retinoic acid-induced differentiation of human leukemia HL-60 cells, *International journal of hematology*, 92 (2010) 136-143.
- [299] M.B. Abubakar, W.Z. Abdullah, S.A. Sulaiman, B.S. Ang, Polyphenols as key players for the antileukaemic effects of propolis, *Evidence-Based Complementary and Alternative Medicine*, 2014 (2014).
- [300] G. Han, J. Xia, J. Gao, Y. Inagaki, W. Tang, N. Kokudo, Anti-tumor effects and cellular mechanisms of resveratrol, *Drug discoveries & therapeutics*, 9 (2015) 1-12.
- [301] A. Rauf, M. Imran, M.S. Butt, M. Nadeem, D.G. Peters, M.S. Mubarak, Resveratrol as an anti-cancer agent: A review, *Critical reviews in food science and nutrition*, 58 (2018) 1428-1447.
- [302] L. Wang, L. Long, W. Wang, Z. Liang, Resveratrol, a potential radiation sensitizer for glioma stem cells both in vitro and in vivo, *Journal of pharmacological sciences*, 129 (2015) 216-225.
- [303] C. Forni, M. Rossi, I. Borromeo, G. Feriotto, G. Platamone, C. Tabolacci, C. Mischiati, S. Beninati, Flavonoids: A myth or a reality for cancer therapy?, *Molecules*, 26 (2021) 3583.
- [304] S. You, M.-J. Wang, Z.-Y. Hou, W.-D. Wang, T.-T. Du, N.-N. Xue, M. Ji, X.-G. Chen, Chlorogenic Acid Induced Neuroblastoma Cells Differentiation via the ACAT1-TPK1-PDH Pathway, *Pharmaceuticals*, 16 (2023) 877.
- [305] X. He, Y. Liao, J. Liu, S. Sun, Research Progress of Natural Small-Molecule Compounds Related to Tumor Differentiation, *Molecules*, 27 (2022) 2128.
- [306] E. Charafe-Jauffret, C. Ginestier, F. Iovino, J. Wicinski, N. Cervera, P. Finetti, M.H. Hur, M.E. Diebel, F. Monville, J. Dutcher, M. Brown, P. Viens, L. Xerri, F. Bertucci, G. Stassi, G. Dontu, D. Birnbaum, M.S. Wicha, Breast cancer cell lines contain functional cancer stem cells with metastatic capacity and a distinct molecular signature, *Cancer Res*, 69 (2009) 1302-1313.
- [307] S.F. S, K. Szczesna, M.S. Iliou, M. Al-Qahtani, A. Mobasher, J. Kobolak, A. Dinnyes, In vitro models of cancer stem cells and clinical applications, *BMC Cancer*, 16 (2016) 738.
- [308] R. Foster, R.J. Buckanovich, B.R. Rueda, Ovarian cancer stem cells: working towards the root of stemness, *Cancer letters*, 338 (2013) 147-157.
- [309] O. Panawan, A. Silsirivanit, C.H. Chang, S. Putthisen, P. Boonnate, T. Yokota, Y. Nishisyama - Ikeda, M. Detarya, K. Sawanyawisuth, W. Kaewkong, Establishment and characterization of a novel cancer stem - like cell of cholangiocarcinoma, *Cancer Science*, (2023).
- [310] M. Kapałczyńska, T. Kolenda, W. Przybyła, Zaj [a with cedilla] czkowska M, Teresiak A, 910-919.
- [311] A. Nyga, U. Cheema, M. Loizidou, 3D tumour models: Novel, in, *Vitro*.
- [312] K. Unnikrishnan, L.V. Thomas, R.M. Ram Kumar, Advancement of scaffold-based 3D cellular models in cancer tissue engineering: an update, *Frontiers in oncology*, (2021) 4468.
- [313] O. Habanjar, M. Diab-Assaf, F. Caldefie-Chezet, L. Delort, 3D cell culture systems: tumor application, advantages, and disadvantages, *International journal of molecular sciences*, 22 (2021) 12200.
- [314] S.-p. Qiao, Y.-f. Zhao, C.-f. Li, Y.-b. Yin, Q.-y. Meng, F.-H. Lin, Y. Liu, X.-l. Hou, K. Guo, X.-b. Chen, An alginate-based platform for cancer stem cell research, *Acta biomaterialia*, 37 (2016) 83-92.
- [315] A. Biddle, In vitro cancer models as an approach to identify targetable developmental phenotypes in cancer stem cells, *In vitro models*, (2023) 1-6.

[316] Y. Tabata, Tissue regeneration based on tissue engineering technology, *Congenital anomalies*, 44 (2004) 111-124.

[317] S. Bandhavkar, Cancer stem cells: a metastasizing menace!, *Cancer medicine*, 5 (2016) 649-655.

[318] G. Hassan, S.M. Afify, S. Kitano, A. Seno, H. Ishii, Y. Shang, M. Matsusaki, M. Seno, Cancer stem cell microenvironment models with biomaterial scaffolds in vitro, *Processes*, 9 (2020) 45.

[319] R. Bartlett, W. Everett, S. Lim, G. Natasha, M. Loizidou, G. Jell, A. Tan, A.M. Seifalian, Personalized in vitro cancer modeling—fantasy or reality?, *Translational oncology*, 7 (2014) 657-664.