

ANALGESIC AND ANTIEMETIC ACTIVITIES OF *FICUS EXASPERATA* VAHL., AND *CLEOME CILIATA* SCHMACH AND THONN.

Misbah Zubair¹, Syed Waseemuddin Ahmed¹, Salman Ahmed^{1*}, M.Shaiq Ali² and Patricia Akpomedaye Onocha³

¹Department of Pharmacognosy, Faculty of Pharmacy, University of Karachi, Karachi-75270, Pakistan.

²H.E.J. Research Institute of Chemistry, International Centre for Chemical & Biological Sciences, University of Karachi, Karachi-75270, Pakistan

³ Natural products/Medicinal Chemistry Unit, Department of Chemistry, University of Ibadan, Ibadan, Oyo State, Nigeria

Article Received on
25 June 2014,

Revised on 20 July 2014,
Accepted on 15 August 2014

***Correspondence for**

Author

Salman Ahmed

Department of Pharmacognosy
Faculty of Pharmacy,
University of Karachi, Karachi
75270, Pakistan.

ABSTRACT

The present investigation is an attempt to evaluate analgesic and antiemetic activities of *Ficus exasperata*; *Cleome ciliata* leaves methanolic extract by using chick emesis model (oral treatment) and acetic acid induced writhing test (intraperitoneal treatment) in mice respectively. The antiemetic activity (150 mg/kg b.w., of extract) was carried out by using chlorpromazine (150mg/kg) as standard antiemetic drug. The analgesic activity (250 mg/kg b.w., of extract) was performed by using aspirin (150mg/kg) as standard analgesic drug. The results showed significant analgesic and antiemetic effects.

KEYWORDS: *Ficus exasperata*; *Cleome ciliata*; antiemetic; analgesic; chick emesis model; acetic acid induced writhing test.

INTRODUCTION

Ficus exasperata Vahl., the sand paper tree (family Moraceae), is a small to medium-sized tree. Leaves are alternate, leathery and ovate-elliptic. Figs are singly or in pairs, orange-red when ripe. It is widely distributed in Africa, Arabian Peninsula, Angola, Ethiopia, India, Mozambique, Sri Lanka, Senegal, Yemen, Zambia and South-eastern Zimbabwe^{1,2}. Various

parts of plant are used as abortifacient, analgesic, antiarthritic, antiarrhythmic, antidysenteric, diuretic, febrifuge and wound healing. *Ficus exasperata* is used against, cutaneous or subcutaneous parasitic infection, edema, leprosy, ophthalmic and oral infections nasopharyngeal afflictions, rheumatism, gout, kidney disorders, hemorrhoids and venereal diseases. Leaves have been used internally to treat anxiety, arthritis, epilepsy, hypertension, rheumatism^{3,4} easing childbirth, treating leprosy⁵, sexually transmitted diseases, gastroenteritis, ophthalmia, cough and hemorrhoid⁶. Whereas their external use is recommended in asthma, bronchitis, chest pain, emphysema, cardiac arrhythmias, tuberculosis^{4,7}, fungal infections, ringworm, rheumatism, jaundice and inflammation of the gums. Stem bark is used for the treatment of abscesses, abnormal enlargement of the spleen, eye ailments, hemorrhoids and stomach-ache. It is locally applied on the body for the treatment of malaria². Roots are used to manage asthma⁸, eye problems, tuberculosis, venereal diseases and to expel worms². Anti-diabetic, hypotensive, antioxidant, anti-inflammatory, antiarthritic, antinociceptive, anticonvulsant, anxiolytic, antiulcer, antipyretic, antimicrobial, uterotonic⁴, pesticidal⁹ anthelmintic¹⁰, anti-dysmenorrhea¹¹, antimalarial¹², diuretic¹³, anti-gastroenteritic¹⁴, molluscicidal¹⁵ activities are reported from leaves. Bark demonstrates anti-inflammatory¹⁶ and antioxidant¹⁷ properties. Flavonoids, glycosides, saponins, steroids and tannins are reported from leaves, stem and root¹⁸. Fruits contain higher proportion of unsaturated fatty acids (linoleic acid and oleic acid) whereas saturated fatty acids detected include stearic acid and palmitic acid¹⁹. The chemical structures of isolated phytochemicals are mention in Figure 1.

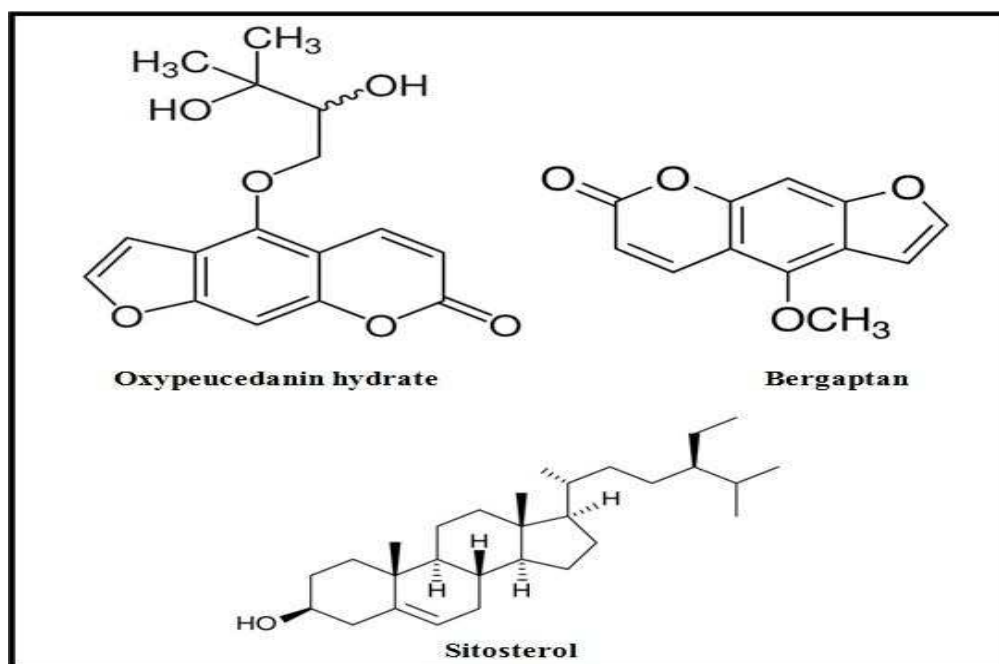


Figure 1: Chemical structure of constituents isolated from *Ficus exasperata* Vahl¹⁶.

Cleome ciliata Schmach and Thonn., the spider plant (family Capparaceae) is a green creeping annual or short-lived perennial herb that spreads like a spider. It is widely distributed in coastal regions of Africa, especially in Nigeria, Uganda, Tanzania and Ghana^{20,21}. The plant grows erect to a height of 30 cm., and then falls flat on the ground where it continues to grow and flower²². It has a slender leaf stalk with trifoliate leaves; the leaflets are net-veined and elliptical with smooth margin²⁰. The flowers are distinctly dimorphous with respect to the ovaries of the mature flower. The normal ovary (fertile) and short ovary (abortive) both types are produced on the same plant²². The mature fruit is a capsule and dry dehiscent. Placement of leaves on the plant is spirally alternate and the fruits arise at the axial of the leaves²⁰. The seeds are attached along the sutures formed by the two carpels of the fruit and average 50 seeds per pod. They are small and reniform with the embryo curled up with the sharp radical end touching the cotyledons to form a disc-like structure²⁰⁻²². The plant juice is used for earache, convulsions²¹ and peptic ulcers²³. Leaves are anthelmintic and carminative²⁴ and their sap is externally applied for chronic otitis media²⁵. The leaves of the plant possess antibacterial properties. Preliminary phytochemical investigation suggests the presence of alkaloids, flavonoids, glycosides, saponins, steroids, tannins and terpenoids²⁶. The present investigation was conducted to evaluate the analgesic and antiemetic activities of *Ficus exasperata* and *Cleome ciliata* leaves extracts. Although the analgesic effect of *Ficus exasperata* leaves extract has been studied earlier by using formalin test²⁷. The presented attempt is further justification of analgesic effect by using acetic acid induced writhing test. Antiemetic effect is reported for the first time.

MATERIALS AND METHODS**Collection of Plant material**

The leaves of *Cleome ciliata* Schmach and Thonn, and *Ficus exasperata* Vahl, were collected from the forest of Ibadan, Nigeria and identified by Mr. Felix Usang of Forest Research Institute of Nigeria (FRIN). Voucher specimen was deposited at the Herbarium of the Institute.

Preparation of the plant extracts

Fresh plant materials (2 kg each) of leaves of *Cleome ciliata* Schmach and Thonn, and *Ficus exasperata* Vahl, were soaked separately in methanol for a week and the extracts were

condensed to dryness by evaporation using rotary evaporator at 40°C. These concentrated methanol extracts were used for bioassay.

Animals

Young male chicks, 4 days old (32-52 g) and Swiss albino mice (weighing 20-30 g) of both sexes were obtained from Big-bird Poultry Breeders (Pvt) Ltd., and Animal house of Aga Khan University, Karachi, Pakistan respectively. They were housed in plastic cages with saw dust as beddings under temperature $25 \pm 2^\circ\text{C}$; 12 h/12 h light-dark cycle and given food and water *ad libitum*. Permission and approval from animal studies were obtained from Board of Advanced Studies and Research, University of Karachi [BASR. Res. No.09(46)-2006]. Chicks (for antiemetic activity) and mice (for analgesic activity) were randomly divided into four groups.

Drugs and Chemicals

Acetic acid, copper sulfate (Scharlau Chemie S.A. Barcelona, Spain), chlorpromazine (ICN, USA), Dimethyl sulfoxide (DMSO), tween 80 and methanol (Merck, Darmstadt, Germany) were used in the experiment.

Acute toxicity test

Acute toxicity studies demonstrate that the leaves of *Ficus exasperata*²⁸ and *Cleome ciliata* are safe for administration²⁶.

Analgesic activity

Mice were randomly divided into four groups of seven animals each. Dose of 250mg/kg of *Cleome ciliata* and *Ficus exasperata* were administered to two groups while the remaining two groups received distilled water 10 ml/kg and aspirin 150 mg/kg respectively, following protocols established by Koster *et al.*, (1959)²⁹ and Salawu *et al.*, (2008)³⁰. All treatments were administered intraperitoneally, and after 30 min 10 ml/kg of 0.6% acetic acid solution in normal saline were injected intraperitoneally. The numbers of writhes were counted for 15 mins after acetic acid injection. The percentage inhibition was calculated using formula³¹:

$$(N - N_t / N) \times 100$$

Where

N = Average number of writhes in control group ; N_t= Average number of writhes in test group

Antiemetic activity

The antiemetic activity was determined by following the protocols of Akita *et al.*, 1998³². Each chick was set aside in a large beaker for 10 minutes to stabilize. Chlorpromazine and extracts were dissolved in 0.9 % saline containing 5 % DMSO and 1 % tween 80 and administered abdominally at a dose of 150 mg/kg b.w., to the test animal. After 10 minutes copper sulfate was administered orally at 50 mg/kg b.w., to each chick, then the number of retching was observed during the next 10 minutes. The percent inhibition was calculated by the following formula:

$$\text{Inhibition (\%)} = [(A-B)/A] \times 100$$

Where

A = Frequency of retching in control group ; B = Frequency of retching in test groups

RESULTS AND DISCUSSION

Ficus exasperata and *Cleome ciliata* significantly decreased the number of writhes when compared to the aspirin treatment and the control (**Table 1 and Figure 2**). *Ficus exasperata* and *Cleome ciliata* in dose of 250 mg/kg body weight reduced the numbers of writhes by 49.78 and 36.87%, respectively. The group of mice treated with aspirin at a dose of 150 mg/kg body weight had 13 writhes compared to the 69.7 writhes of the control group, thus aspirin reduced the writhes by 81.34%. The acetic acid induced writhing method is an effective method to evaluate peripherally active analgesics. The abdominal constriction response induced by acetic acid is a sensitive method to test peripherally acting analgesics³³. Hyperalgesia, induced by the injection of acetic acid, is characterized by contraction of the abdominal muscle accompanied by body elongation and an extension of the forelimbs. Various peripherally acting analgesic drugs such as ibuprofen, aspirin and indomethacin have been reported to inhibit acetic acid induced writhing in mice^{33,34}. Tested extracts of *Ficus exasperata* and *Cleome ciliata* leaves reported effective against acetic acid induced writhing in mice and has been suggested to be a peripherally acting analgesic³⁵, perhaps via the inhibition of synthesis and release of prostaglandins and other endogenous substances³⁰. The mechanism of action may be linked to the inhibition of cyclooxygenases.

Table 1: Effect of *Ficus exasperata* and *Cleome ciliata* on acetic acid induced writhing in mice.

| Treatments | Dose (mg/kg i.p.,) | Mean Number of writhes ± S.E.M. (15 mins) | % age inhibition of writhes |
|------------|--------------------|---|-----------------------------|
| Control | ----- | 69.7±3.44 | ----- |
| AS | 150 | 13.0±2.1 | 81.34 |

| | | | |
|----|-----|-----------|-------|
| FE | 250 | 35.0±1.3 | 49.78 |
| CC | 250 | 44.0±0.57 | 36.87 |

AS= Aspirin ; FE= *Ficus exasperata* ; CC= *Cleome ciliata* ; S.E.M = Standard Error of Mean, N=7, * $p < 0.05$ vs. control applying unpaired t -test.

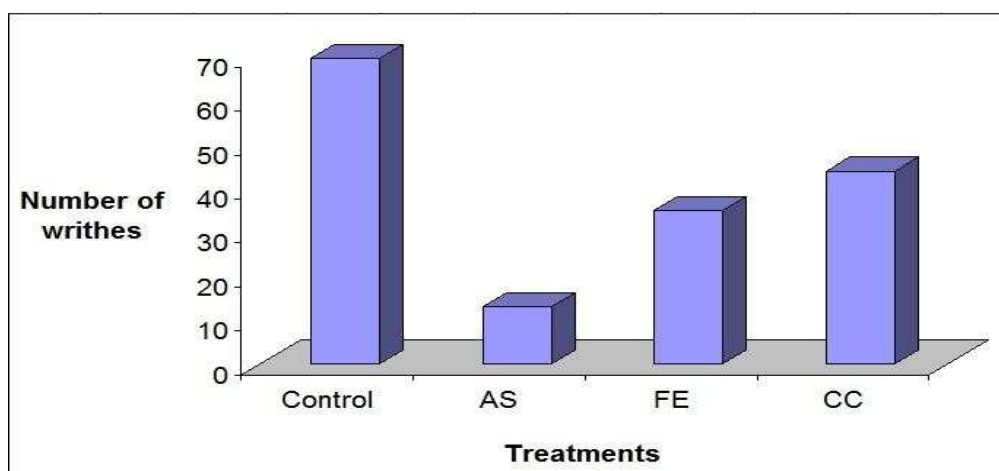


Figure 2: Analgesic effect of *Ficus exasperata* and *Cleome ciliata*

The results of antiemetic effect of methanol extracts of *Ficus exasperata* and *Cleome ciliata* are shown in **table 2 and figure 3**. Leaves extracts of *Ficus exasperata* and *Cleome ciliata* showed 46.21% and 58.20% inhibition of retches. The standard drug chlorpromazine showed 35.23 % inhibition of retches. The methanol extracts of *Ficus exasperata* and *Cleome ciliata* showed significant ($p < 0.05$) anti-emetic effect in young chicks. The protective effect of the extract against copper sulfate induced retching in young chicks is possibly by peripheral action as the oral copper sulfate induces emesis by peripheral action through excitation of visceral afferent nerve fibers of the GIT³⁶. It has also been established that the peripheral 5-HT₃³⁷, 5-HT₄³⁸ or NK₁³⁹ receptors are involved in emesis. Therefore, it may be said that the methanol extracts of *Ficus exasperata* and *Cleome ciliata* produced antiemetic activity by receptor antagonism and has peripheral antiemetic action. Moreover, anti-emetic activity by using copper sulfate proposed 5-HT₃³⁷, 5-HT₄³⁸ or NK₁³⁹ receptors antagonism. Therefore it may be said that extracts were able to effectively prevent emesis and has a peripheral antiemetic action.

Table 2: The anti-emetic effect of *Ficus exasperata* and *Cleome ciliata* on copper sulfate induced emesis in chicks.

| Treatments | Number of Retches (Mean ± SEM) | Inhibition (%) of retches |
|------------|--------------------------------|---------------------------|
| Control | 70.10 ± 3.28 | ----- |
| CZ | 45.40 ± 3.11* | 35.23 |
| FE | 37.70 ± 3.12* | 46.21 |

| | | |
|----|-------------|-------|
| CC | 29.30±1.92* | 58.20 |
|----|-------------|-------|

CZ = Chlorpromazine; FE= *Ficus exasperata* ; CC= *Cleome ciliata* ; S.E.M = Standard Error of Mean, N=7, Dose=150 mg/kg p.o., *= $p < 0.05$ significantly different from control value using unpaired student's *t*-test.

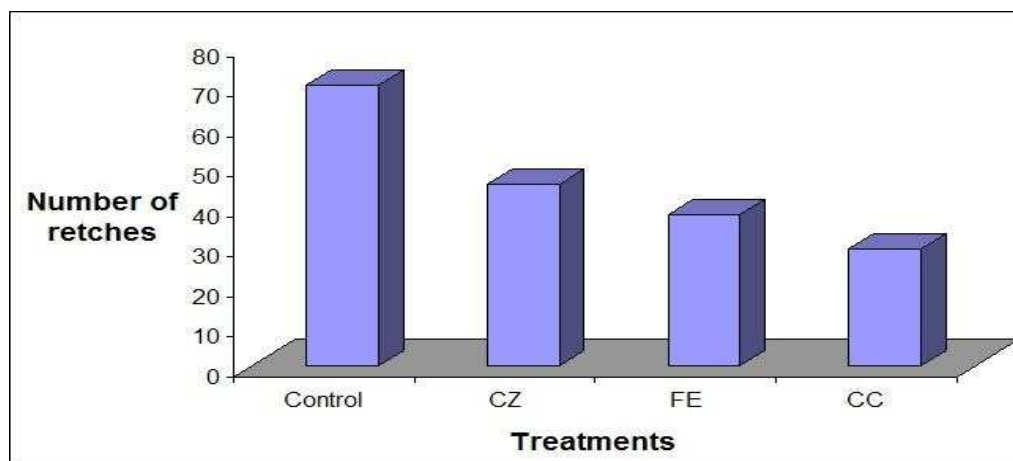


Figure 3: Antiemetic effect of *Ficus exasperata* and *Cleome ciliata*

As mentioned earlier that flavonoids, steroids and terpenoids are reported from *Ficus exasperata*¹⁸ and *Cleome ciliata*²⁶. Flavonoids, steroids and terpenoids have analgesic and antiemetic activities⁴⁰. So, it may be implied that the observed analgesic and antiemetic effects were due to the presence of these phytochemicals. However compound targeted activities are further required to justify the responsible analgesic and antiemetic compound(s) and other bioassays to clarify actual mechanism.

REFERENCES

1. Flora of Zimbabwe. *Ficus exasperata* Vahl., 2009; http://www.zimbabweflora.co.zw/speciesdata/species.php?species_id=120280
2. Niangadouma R. *Ficus exasperata* Vahl. In: Brink, M. & Achigan-Dako, E.G. (Editors). Prota 16: Fibres/Plantes à fibres. PROTA, Wageningen, Netherlands. 2010. http://database.prota.org/PROTAhtml/Ficus%20exasperata_En.htm
3. Ahmed F, Mueen KK, Zainul Abedin Md, Karim AA. Traditional uses and pharmacological potential of *Ficus exasperata* Vahl. *Sys Rev Pharm*, 2012; 3(1): 15-23.
4. Irvine FR. *Woody plants of Ghana*. 1st Edition, Oxford University Press, London. 1961.
5. Burkill HM. *The useful plants of tropical West Africa*. 43rd Ed. Royal Botanic Gardens Kew, London, UK., 1997.
6. Borokini TI and Omotayo FO. Comparative Phytochemical Analysis of selected Medicinal Plants in Nigeria. *Int J Advanced Chem Res*, 2012; 1(1):011-018.

7. Assi AL. Utilisation de diverses espèces de Ficus (Moraceae) dans la pharmacopée traditionnelle africaine de Côte d'Ivoire. *Mitteilungen aus dem Institut für allgemeine Botanik in Hamburg*, 1990; 23:1039–1046.
8. Chhabra CE, Mahunnah RLA, Mshio EN. Plants used in traditional medicine in Eastern Tanzania IV Angiosperms (Mimosaceae to Papilionaceae). *J Ethnopharmacol.* 1990; 29:295-323.
9. Igoli JO, Ogaji OG, Tor-Anyiin TA, Igoli NP. Traditional medicine practice amongst the Igede people of Nigeria Part II. *Afr J Trad Compl Med.* 2005; 2:134–152.
10. Nweze NE, Ogidi A, Ngongeh LA. Anthelmintic potential of three plants used in Nigerian ethnoveterinary medicine. *Pharm Biol.* 2013; 51(3):311-5.
11. Bafor EE, Omogbai EI, Ozolua RI. Oxytocin inhibiting effect of the aqueous leaf extract of *Ficus exasperata* (Moraceae) on the isolated rat uterus. *Act Poloniae Pharmaceutica-Drug Res*, 2011; 68(4):541-7.
12. Okon OE, Gboeloh LB, Udoh S. Anti-plasmodial effect of *Ficus exasperata* on albino mice experimentally infected with *Plasmodium berghei* (NK65). *Universal J Pharm.* 2013; 2(5):29-35.
13. Amonkan AK, Konan AB, Ahui BM, Bleyéré MN, Kouakou LK, Bouafou GM. Diuretic effects of aqueous extract of *Ficus exasperata* Vahl. leaves in rat. *Pak J Bio Sci.* 2013; 16(21):1383-7.
14. Chifundera K. Livestock diseases and the traditional medicine in the bushi area, Kivu province, Democratic Republic of Congo. *Afr Study Monographs*, 1998; 19(1):13-33.
15. Oledibe PA, Morenikeji OA, Benson O. Molluscicidal potency of *Ficus exasperata* (Vahl) against juvenile and adult *Biomphalaria pfeifferi*. *Zoology & Ecol.* 2013; 23(2):147–156.
16. Amponsah IK, Fleischer TC, Dickson RA, Annan K, Thoss V. Evaluation of anti-inflammatory and antioxidant activity of Furanocoumarins and Sterolin from the stem bark of *Ficus exasperata* Vahl (Moraceae). *J Sci & Innov Res.* 2013; 2(5):880-7.
17. Mouho D, Ouattara Z, Zabri H. Anti-radical potentiality of the extract methanolique of the bark of a Plant used in traditional medicine in Cote D'ivoire : *Ficus exasperata* (Moraceae). *J Appl Chem*, 2014; 2(2):65-71.
18. Adebayo EA, Ishola OR, Taiwo OS, Majolagbe ON, Adekeye BT. Evaluations of the methanol extract of *Ficus exasperata* stem bark, leaf and root for phytochemical analysis and antimicrobial activities. *Afr J Plant Sci.* 2009; 3(12):283-7.

19. Bello MO, Abdul-Hammed M, Adepoju AJ, Esan OA, Tihamiyu AA. Nutritional Composition and fatty acids profile of *Ficus exasperata* fruit and fruit oil. J Nat Sci Res. 2014; 4(2):25-9.
20. Oliver EWHM. Medicinal plants of Nigeria, Part II. Technical Memorandum No. 7, Federal Institute of Industrial Research. Published by the Federal Ministry of Commerce and Industry, Lagos, Nigeria, 1959.
21. Grubben GJH, Denton OA. *Vegetables*. PROTA Foundation, The Netherlands, 2004.
22. Johnson A, Seng TK. *Cleome ciliata* Schum. et Thonn. in Singapore. *Gardens Bulletin* 1958; Vol. XVII. pp.325-330.
23. ASICUMPON. The Association for Scientific Identification, Conservation and Utilization of Medicinal Plants of Nigeria Checklist of Medicinal Plants of Nigeria and their uses. Trinity – Biz Publishers, Abakpa-Enugu, 2005.
24. Gills LS. *Ethnomedical uses of plants in Nigeria*. University of Benin press, Nigeria, 1992; p.276.
25. Utsalo SJ, Onoyom-Ita V, Ifeanyi-Chukwu M, Akpan JO. Home medication and microbiological profile in chronic otitis media in some Nigerian children. The Central Afr J Med, 1990; 36(11):278-283.
26. Umerie SC, Okorie NH, Ezea SC, Okpalaononuju AN. Antibacterial screening and phytochemical analysis of *Cleome ciliata* (Capparidaceae) leaves. Int J Current Res Rev, 2012; 4(15):5-10.
27. Woode E, Poku RA, Abotsi WKM. Anticonvulsant effect of a leaf extract of *Ficus exasperata* Vahl (Moraceae) in mice. Int J Pharmacol. 2011; 1:1-5.
28. Bafor EE, Igbinuwen O. Acute toxicity studies of the leaf extract of *Ficus exasperata* on haematological parameters, body weight and body temperature. J Ethnopharmacol. 2009; 123: 302–307.
29. Koster RM, Anderson, De Beer EJ. Acetic acid for analgesic screening. Fed Proc. 1959; 18: 412.
30. Salawu OA, Chindo BA, Tijani AY, Adzu B. Analgesic, anti-inflammatory, antipyretic and antiplasmodial effects of the methanolic extract of *Crossopteryx febrifuga*. J Med Plants Res. 2008; 2(9): 213-218.
31. Sulaiman MR, Somchit MN, Israf DA, Ahmad Z, Moin S. Antinociceptive effect of *Melastoma malabathricum* ethanolic extract in mice. Fitoter. 2004; 75(7-8): 667-72.
32. Akita Y, Yang Y, Kawai T, Kinoshita K, Koyama K, Takahashi K. New assay method for surveying anti-emetic compounds from natural sources. Nat Prod Sci. 1998; 4:72-77.

33. Gene RM, Segura L, Adzet T, Marin E, Iglesias J. *Heterotheca inuloides*: Anti-inflammatory and analgesic effects. J Ethnopharmacol. 1998; 60: 157-162.
34. Okpo SO, Fatokun F, Adeyemi OO. Analgesic and anti-inflammatory activity of *Crinum glaucum* aqueous extracts. J Ethnopharmacol. 2001; 78: 207-211.
35. Singh S, Majumdar DK. Analgesic activity of *Ocimum sanctum* and its possible mechanism of action. Int J Pharmacog. 1995; 33: 188.
36. Bowman WC, Rand MJ. *Textbook of pharmacology*, Oxford, Blackwell Scientific Publication, 1980.
37. Fukui H, Yamamoto M, Sasaki S, Sato S. Involvement of 5-HT₃ receptors and vagal afferents in copper sulfate- and cisplatin-induced emesis in monkeys. Euro J Pharmacol. 1993; 249:13-18.
38. Fukui H, Yamamoto M, Sasaki S, Sato S. (1994). Possible involvement of peripheral 5-HT₄ receptors in copper sulfate-induced vomiting in dogs. Euro J Pharmacol.1994; 257:47-52.
39. Ariumi H, Saito R, Nago S, Hyakusoku M, Takano Y, Kamiya H-O. The role of tachykinin NK1 receptors in the area postrema of ferrets in emesis. Neurosci Lett. 2000; 286:123-126.
40. Shaheen N, Ahmed S, Azhar I, Hasan MM. Analgesic, anti-inflammatory and antiemetic activities of *Cleome scaposa* DC. Phytopharmacol. 2013; 4(1), 106-113.