ANTIFUNGAL ACTIVITY OF HYPNEA PANNOSA J. AGARDH

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ABSTRACT

Hypnea pannosa J. Agardh (methanol extract of the whole alga) was subjected to antifungal screening. Extract showed good activity against *Trichophyton longifusus*, low activity against *Candida glabrata* and *Microsporum canis*. The extract was found to be inactive against *Fusarium solani*.

Keywords: Candida, Hypnea, Microsporum, Trichophyton, Antifungal Activity, Red Algae, Pakistan.

INTRODUCTION

Hypnea pannosa J. Agardh is a red marine alga, belongs to family Hypneaceae (Valeem & Shameel 2012). H. pannosa is distributed in Atlantic Islands: (Cape Verde Islands), North America: (Gulf of California and Mexico), Central America: (Costa Rica, El Salvador, México-Pacific), Nicaragua, Panama; South America: Brazil, Galápagos Islands; Africa: Eritrea, Madagascar, Mauritius, Senegal, Tanzania; Indian Ocean Islands: Aldabra Islands, Chagos Archipelago, Christmas Island, Diego Garcia Atoll, Laccadive Islands, Maldives, Réunion, Seychelles; South-west Asia: Bangladesh, India, Iran, Oman, Pakistan, Sri Lanka, Yemen; Asia: China, Japan, Korea, Taiwan; South-east Asia: Indonesia, Philippines, Singapore, Thailand, Vietnam; Australia and New Zealand: Queensland, Western Australia; Pacific Islands: Federated States of Micronesia, Fiji, French Polynesia, Hawaiian Islands, Mariana Islands, Samoa, Samoan Archipelago, Solomon Islands (Guiry & Guiry 2013). Amino acid such as alanine, arginine, aspartic acid, glutamic acid, glycine, isoleucine, leucine, lysine, valine (Siddique et al. 2013) and brominated sesquiterpenes as 10-bromo-7, 12-dihydroxy- $\Delta^{3,4}$ -laurene, filiformin and filiforminol (Fig. 1) were the reported chemical constituents (Afaq-Hussain et al. 1991). H. pannosa possessed antibacterial (Shanmughapriya et al. 2008), analgesic, anti-emetic (Mazhar et al. 2011) and haemagglutinic activity (Alam & Usmanghani 1994).

Although the antifungal activity of *H. pannosa* against *Candida albicans* had been reported earlier (Shanmughapriya *et al.* 2008). Present study is the continuation of antifungal activity against *Candida glabrata*, *Fusarium solani*, *Microsporum canis* and *Trichophyton longifusus*.

MATERIALS & METHODS

Collection of plant materials

Hypnea pannosa was collected from Manora and Buleji the coastal areas of Karachi and dried under shade. Sample was deposited in the herbarium of the Department of Pharmacognosy, Faculty of Pharmacy, University of Karachi for further reference.

Preparation of the extracts

The dried alga was crushed and soaked in methanol for seven days. The methanol extract was evaporated under reduced pressure at 35° C, following the protocol of Rizvi & Shameel (2005).

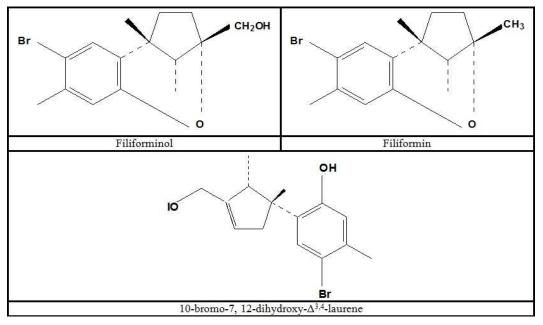


Fig. 1. Structure of constituents isolated from Hypnea pannosa (Afaq-Hussain et al., 1991).

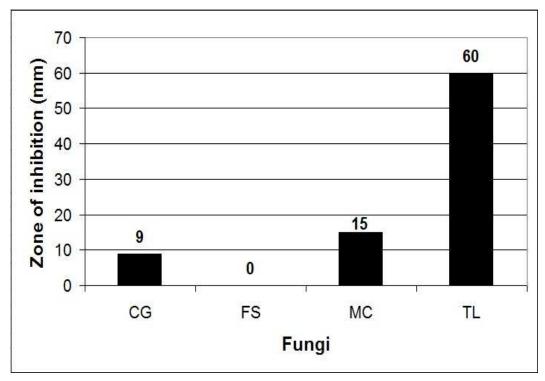


Fig. 2. Antifungal Activity of *Hypnea pannosa*. (CG= *Candida glabrata*, FS = *Fusarium solani*, MS= *Microsporum canis* and TS= *Trichophyton longifusus*).

	Linear growth (mm)		Zone of	Std. Drug
Fungi	Control	Methanol Extract	inhibition (mm) methanol extract 400 µg/mL	(Miconazole) MIC µg/mL
Candida glabrata	100	91	9	110.8
Fusarium solani	100	100	0	73.25
Microsporum canis	100	85	15	98.4
Trichophyton longifusus	100	40	60	70

Table I. Antifungal Activity	y of <i>nyphea pannosa</i>
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Antifungal Activity

The antifungal activity of methanol extract of *Hypnea pannosa* was performed using tube dilution method (Washington & Sutter 1980). Sabouraud dextrose agar was prepared by mixing 32.5 g sabouraud 4 % glucose and 4.0 g of agar-agar in 500 mL distilled water. It was steamed to dissolve and 4 mL was dispensed in screw capped tubes and autoclaved at 121° C for 15 min. Tubes were allowed to cool to 50° C. Stock solution of the extract was prepared by dissolving 24 mg in 1 mL DMSO. The stock solution was added to solidified sabouraud agar media to give 400 μ g crude extract/mL of sabouraud dextrose agar. Tubes were allowed to solidify in slanting position and inoculated with 4 mm diameter piece of the incolum removed from 7 days old culture of fungi. For non-mycelial growth, an agar surface streak was employed. DMSO and reference antifungal drugs were served as negative and positive control, respectively. The test tubes were incubated at 27-29° C for 7-10 days. Growth in the medium containing the extract was determined by measuring linear growth (mm) and zone of growth inhibition (mm) was calculated with reference to negative control. In that assay, Miconazole was used as standard antifungal drug.

RESULTS & DISCUSSION

Plants have been a source of medicinal compounds since pre-historic time. It has been a well established fact that all parts of plants are used in traditional system of medicine for centuries. Organic compounds from terrestrial and marine organisms had extensive past and present use in the treatment of many diseases and served as compounds of interest both in their natural form and templates for synthetic modification (Chin *et al.* 2006).

The antifungal activity of the methanol extract of the *Hypnea pannosa* (400 μ g/mL) was carried out by tube dilution method. The antifungal activity was presented in Table I and Fig. 2. The methanol extract of *Hypnea pannosa* showed good activity against *Trichophyton longifusus* and low activity against *Candida glabrata* and *Microsporum canis*, whereas it was inactive against *Fusasium solani*. Brominated sesquiterpenes from red alga reported to possess anti fungal activity (Ji *et al.* 2007). Brominated sesquiterpenes was also reported from *Hypnea pannosa* (Afaq-Hussain *et al.* 1991). Therefore, it may be concluded that these sesquiterpenes play some role in anti fungal effect of *Hypnea pannosa*.

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