



ANALGESIC ACTIVITY OF LEAVES EXTRACTS OF *SAMANEA SAMAN* MERR., AND *PROSOPIS CINERARIA* DRUCE.

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ABSTRACT

Current study was designed to explore the analgesic effects of methanol extracts of the leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce., using tail immersion test. The painful reactions in mice were produced by thermal stimuli through dipping the tail tips of mice into hot water. Methanol extracts of the leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce., were administered intraperitoneally at the dose of 100mg /kg body weight. Pethidine 50mg/Kg intraperitoneally was used as standard analgesic drug. The tail flick latency delay was measured at 0.5, 1.0, 1.5, 2.0, 2.5 and 3.0 hour after the intraperitoneal administration. Both extracts produce analgesic effects when compare with pethidine.

KEYWORDS: Analgesic activity, *Samanea saman* Merr., *Prosopis cineraria* Druce., tail immersion test.

INTRODUCTION

Samanea saman Merr., and *Prosopis cineraria* Druce., are medicinal plants of Pakistan and India belong to the family Mimosaceae¹. *Samanea saman* Merr., is a folk remedy for colds, diarrhoea, headache, intestinal ailments, and stomach ache². Leaves are used in diarrhea³. Inner bark decoction is used for the treatment of colds and diarrhea³. Seeds of *Samanea saman* Merr., are chewed for sore throat^{4,5}. Antifungal⁶ and antioxidant⁷ activities are reported from *Samanea saman* Merr. Leaves possess antiemetic⁸, antibacterial⁹ and insecticidal¹⁰ activities. Bark possesses hepatoprotective activity¹¹. Pods possess antibacterial and antifungal activity¹². Alkaloids, glycosides and terpenes are reported from *Samanea saman* Merr¹³. Leaves contain flavonoids, glycosides, saponins, steriods, tannins, and terpenoids¹⁴. Pods indicated the presence of alkaloids, flavonoids, saponins, steroids and tannins¹².

The leaves of *Prosopis cineraria* Druce., are used in cataract, dyspepsia, earache and toothache. The stem bark possesses abortifacient and laxative properties. It is used to treat anxiety, asthma, bronchitis, fever, dysentery, dyspepsia¹⁵ and rheumatism¹⁶. Flowers are used as an anti-diabetic agent and to prevent abortion¹⁷. Root is antidysenteric¹⁶. *Prosopis cineraria* Druce., possesses antitumor activity¹⁸. Antibacterial¹⁹, anthelmintic²⁰, antipyretic²¹, antioxidant, hypoglycemic and hypolipidemic activities²² are reported from stem bark. Literature survey of *Prosopis cineraria* Druce., revealed the presence of alkaloids²³, fatty acids²⁴, glycosides and sterols²⁵ whereas glucosides are reported from flowers and flavones from seeds¹⁶.

The tail immersion test is used for evaluating central antinociceptive activity by responding to the pain stimuli conducting through neuronal pathways²⁶. The purpose of the present study is to investigate analgesic activity of the leaves (methanol extracts) of *Samanea saman* Merr., and *Prosopis cineraria* Druce., in Swiss albino mice using tail immersion test. The stem bark²⁴ and roots²⁷ of *Prosopis cineraria* Druce., are reported to possess analgesic activity. The analgesic potential of leaves (aqueous extract) by acetic acid induced writhing test has already studied²⁸. Here we evaluate methanol extract by using tail immersion test to further confirm its central analgesic effect. The analgesic effect of *Samanea saman* Merr., is reported first time.

MATERIALS AND METHODS

Plant Sample Collection and Identification

Leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce., were collected in summer 2012 from Karachi, Pakistan and compared with already deposited voucher specimen of *Samanea saman* Merr., (K-97-13) and *Prosopis cineraria* Druce.,(K-97-05).

Plant Extraction

Leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce., were dried under shade and soaked in methanol for a week. The extracts were filtered then concentrated using rotary evaporator at 40°C.

Animals

Male Swiss albino mice (17–23 g) were obtained from the Animal house of Aga Khan University and hospital, Karachi, Pakistan. During the acclimatization period (1 week), the animals were supplied with a standard commercial diet and water *ad libitum* and kept in room temperature. The experimental procedures were carried out in accordance with the ethical guidelines for investigations of experimental pain in conscious animals given by Zimmermann (1983)²⁹. All mice were equally divided into four groups of seven mice each and transferred into different cages with their identification mark. The first group received subcutaneous 0.9% saline, second group received pethidine (50mg/kg i.p.) as standard analgesic drug whereas remaining two groups treated with methanol extracts of leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce., (100 mg/kg i.p. each).

Acute Systemic Toxicity Test

The acute systemic toxicity of methanol extracts of leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce., in Swiss albino mice suggested that 100mg/kg body weight of each extract is safe for intraperitoneal administration³⁰.

Analgesic Activity

The central analgesic activity of *Samanea saman* Merr., and *Prosopis cineraria* Druce., were evaluated by tail immersion test. This test was performed according to the technique of Janssen *et al.*, (1963)³¹ which later on adapted by Ramabadrans *et al.*, (1989)³².

The lower two-third of the tail was marked and immersed in a water bath having temperature of 55±0.5°C. The time in seconds until the tail was withdrawn from the water was defined as the reaction time. The reaction time was measured

at 0.5, 1.0, 1.5, 2.0, 2.5 and 3.0 hour after the intraperitoneal (i.p.) administration of vehicle, test group (*Samanea saman* Merr., and *Prosopis cineraria* Druce., at the dose of 100mg/kg body weight) and standard (pethidine, 50mg/kg), with the reaction time of zero minutes being the start of the test. While measurements were being made, animals were immobilized. Inhibition of pain (%) or pain threshold was calculated as follows:

$$\text{Pain threshold} = (\text{Treated mean} - \text{Control mean} / \text{Control mean}) \times 100$$

Statistical Analysis

Analgesic activity was expressed as mean \pm standard error of mean. The statistical significance of the difference was determined by an unpaired Student's *t*-test. The values $P < 0.01$ and $P < 0.05$ are statistically significant and more significant vs. control.

Table: Effects of the methanolic extracts of *Samanea saman* and *Prosopis cineraria* on pain threshold of mice in tail immersion test

Treatment	Post treatment time (seconds) TFLD \pm SEM (% inhibition of pain)						Average % analgesia
	0.5hr	1hr	1.5hr	2hr	2.5hr	3hr	
Control	1.20 \pm 0.2	1.0 \pm 0.3	1.60 \pm 0.4	1.20 \pm 0.2	1.10 \pm 0.4	1.2 \pm 0.3	-----
Pethidine (50mg/kg i.p.)	22.52 \pm 0.13** (94.67)	22.57 \pm 0.23** (95.56)	23.52 \pm 0.46** (93.19)	22.2 \pm 0.63** (94.59)	21.92 \pm 0.07** (94.98)	21.57 \pm 0.20* (94.43)	94.57
SS (100mg/kg i.p.)	23.4 \pm 0.68 (94.87)	23.9 \pm 0.68 (95.81)	24.58 \pm 0.03 (93.49)	28.69 \pm 0.70** (95.81)	25.2 \pm 0.60* (95.63)	22.25 \pm 0.70** (94.60)	95.03
PC (100mg/kg i.p.)	24 \pm 0.23 (95.0)	28.12 \pm 0.25 (96.44)	28.9 \pm 0.23 (94.46)	30.2 \pm 0.61** (96.02)	27.2 \pm 0.23 (95.95)	20.36 \pm 0.20 (94.10)	95.32

PC= *Prosopis cineraria*, SS= *Samanea saman*; Tail withdrawal reflex in seconds; TFLD = Tail Flick Latency Difference, n=7 for each group, * $p < 0.01$ and ** $p < 0.05$ are statistically significant and more significant vs. control followed by unpaired students' *t*-test.

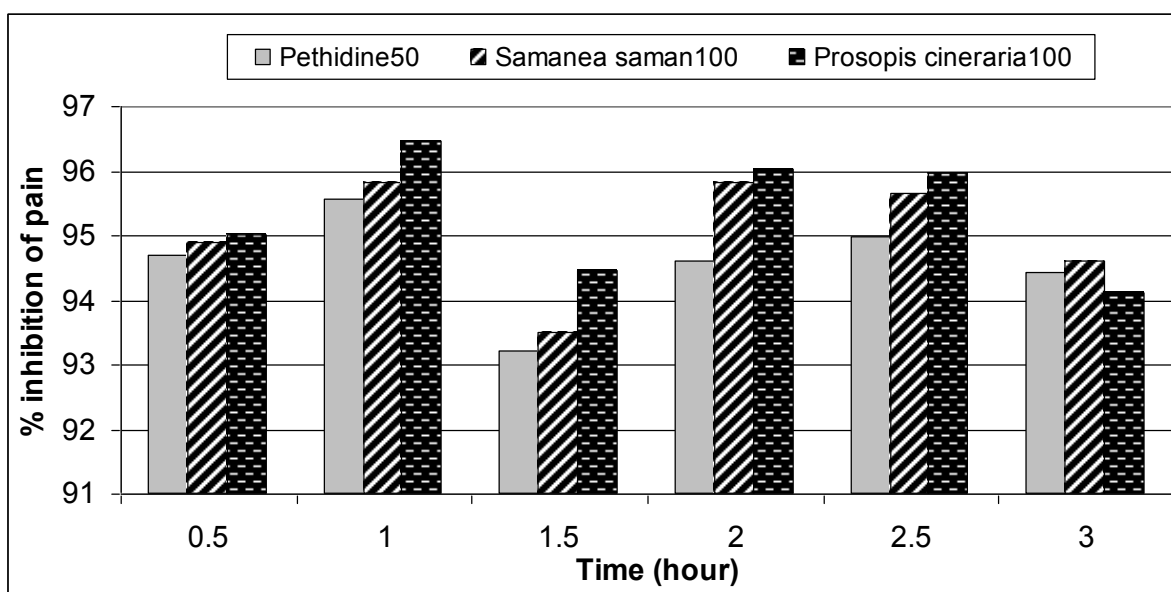


Figure: Analgesic effect of *Samanea saman* and *Prosopis cineraria*.

RESULTS AND DISCUSSION

The methanol extracts of *Samanea saman* Merr., and *Prosopis cineraria* Druce., inhibited tail flick response at 0.5hr in mice. This tail flick latency delay was increased in linear fashion till 1hr and then decreased (table). The inhibitory effects of the extract became pronounced between 0.5 and 2 h post-dosing and reached a maximum of 28.69 sec and 30.2 sec ($p < 0.05$) in case of *Samanea saman* Merr., and *Prosopis cineraria* Druce., respectively. In case of pethidine tail flick latency delay was increased till 1hr and afterward decreases. The leaves extract of *Samanea saman* Merr., and *Prosopis cineraria* Druce., in a dose of 100 mg/kg showed anti-nociceptive activity when compared with pethidine (figure).

The tail immersion test is used for evaluating centrally acting analgesics³³ and is more sensitive to opioid receptor agonists³⁴. This test consists of a thermal stimulus and increase in the reaction time is used for evaluating central antinociceptive activity³⁵. The tail flick response is believed to be a spinally mediated reflex²⁷. So, it differentiates between central and peripheral analgesics³⁶. Opioid agents exhibit their analgesic effects both via supraspinal (μ_1 , κ_3 , δ_1 ,

σ_2) and spinal (μ_2 , κ_1 , δ_2) receptors^{37,38}. Pethidine produces analgesia by stimulating μ (μ), δ (δ) and κ (κ) opioid receptors present in spinal cord and brain stem³⁹. As this test model is for evaluating centrally acting analgesic effect so, it may be said that the methanol extracts of leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce., possess central analgesic effect. However, more centrally acting analgesic models are further needed to confirm this effect.

One possible reason of this central analgesic effect may be the presence of alkaloids and terpenes⁴⁰ already reported in leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce.

The present investigation suggested the central analgesic effects of leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce. However, further studies are required to obtain effective compound(s) from the methanolic extracts of the leaves of *Samanea saman* Merr., and *Prosopis cineraria* Druce., and clarify the possible mechanism of action. The exact mechanism and the bioactive principles responsible for these actions remain to be explained.

REFERENCES

1. Ali SI. Flora of West Pakistan. No.36, Department of Botany:University of Karachi: Karachi, 1972.
2. Duke JA, Wain KK. "Medicinal plants of the world", 1981, 3.
3. Staples GW, Elevitch CR. *Samanea saman* (rain tree) Fabaceae (legume family). Species Profiles for Pacific Island Agroforestry, 2006; Available from: <http://agroforestry.net/tti/Samanea-raintree.pdf>
4. Perry LM. Medicinal Plants of East and South East Asia, MIT Press, Cambridge. 1980; p.231.
5. Ayensu ES. Medicinal Plants of West Indies, Inc. Algonac, M.I. 1981; p.282.
6. Thippeswamy S, Mohana DC, Manjunath K. Screening of *in vitro* antifungal activity of some indian medicinal plants against *Candida albicans* and *Cryptococcus neoformans*. Int J Curr Res 2012; 4(3):37-42.
7. Arulpriya P, Lalitha P, Hemalatha S. *In vitro* antioxidant testing of the extracts of *Samanea saman* (Jacq.)Merr. Der Chemica Sinica 2010; 1(2): 73-79.
8. Ahmed S, Sabzwari T, Hasan MM, Azhar I. Antiemetic activity of leaves extracts of five Leguminous plants. Int J Res Ayur & Pharm 2012; 3(2):251-253.
9. Jagessar RC, Mars A, Gomathinayagam S. Selective Antimicrobial properties of Leaf extract of *Samanea Saman* against *Candida albicans*, *Staphylococcus aureus* and *Escherichia coli* using several microbial methods. J American Sci 2011; 7(3): 108-119.
10. Azhar I, Hasan MM, Mazhar F, Ali MS. Some biological evaluations on *Samanea saman*. Pak J Pharmacol 2009; 26(1): 47-53.
11. Sindhan V, Senthil Velan S, Rakesh Joshi. Hepatoprotective activity of *Samanea saman* (Jacq) Merr bark against CCl₄ induced hepatic damage in albino rats. Int J Institutional Pharm & Life Sci 2012; 2(5): 38- 43.
12. Ukoha P, Cemaluk EAC, Nnamdi OL, Madus EP. Tannins and other phytochemical of the *Samanea saman* pods and their antimicrobial activities. African Journal of Pure and Applied Chemistry 2011; 5(8): 237-244.
13. Nigum SK, Misra G, Mitra CR. Constituents of *Samanea saman* bark. Phytochem 1971; 10(88): 1954-1955.
14. Prasad RN, Viswanathan S, Devi JR, Nayak V, Swetha VC, Archana BR, Parathasarathy N, Rajkumar J. Preliminary phytochemical screening and antimicrobial activity of *Samanea saman*. J Med Plants Res 2008;2(10): 268-270.
15. Kirtikar KR, Basu BD. Indian medicinal plants. Vol. II. International Book Distributors, Dehradun, India, 1984; p. 910.
16. Ukani MD, Limbani NB, Mehta NK. A Review on the Ayurvedic Herb *Prosopis cineraria* (L.) Druce. Ancient Science of Life. 2000;Vol. No XX (1&2):1-13.
17. Khare CP. Indian Medicinal Plants. Springer-Verlag, Berlin/Heidelberg, 2007; p. 518.
18. Robertson S, Narayanan N, Kapoor BR. Antitumor activity of *Prosopis cineraria* (L.) Druce against Ehrlich ascites carcinoma-induced mice. Nat Prod Res 2011; 25: 857–862.
19. Velmurugan V, Arunachalam G, Ravichandran V. Antibacterial activity of stem bark of *Prosopis cineraria* (Linn.) Druce. Archives of Applied Science Research 2010; 2, 147-150.
20. Velmurugan V, Arunachalam G, Ravichandran V. Anthelmintic potential of *Prosopis cineraria* (Linn.) druce stem barks. Asian Journal of Plant Science and Research, 2011; 1 (2): 88-91.
21. Manikandar RVM, Rajesh V, Kumar RS, Perumal P, Raj CD. Analgesic and anti-pyretic activity of stem bark of *Prosopis cineraria* (Linn) Druce. J Pharm Res 2009;2(4):660-662.
22. Sharma N, Garg V, Paul A. Antihyperglycemic, antihyperlipidemic and antioxidative potential of *Prosopis cineraria* bark. Indian J Biochem 2010; 25: 193-200.
23. Rastogi R.P. and B.N. Mehrotra. Compendium of Indian Medicinal Plants: A CDRI Series. (Vol. IV) Lucknow: Publication and Information Directorate, New Delhi, 1995.
24. Gangal S., S. Sharma and A. Rauf, Fatty Acid Composition of *Prosopis cineraria* Seeds. Chem Nat Compds 2009; 45(5): 705 – 707.
25. Akhtar H, Virmani OP. Dictionary of Indian Medicinal Plants. (1st Ed) (Central Institute of Medicinal and Aromatic Plants: Lucknow, 1992).
26. Chapman CR, Casey KL, Dubner R, Foley KM, Gracely RH, Reading AE. Pain measurement: an overview. Pain 1985; 22:1–31.
27. Kumar A, Yadav SK, Singh S, Pandeya SN. Analgesic activity of ethanolic extract of roots of *Prosopis cineraria* (L.) Druce. J Appl Pharm Sci 2011; 01 (08): 158-160.
28. Joseph L, George M, Sharma A, Gopal N. Antipyretic and analgesic effects of the aqueous extract of the *Prosopis cineraria*. Global J Pharmacol 2011; 5(2): 73-77.
29. Zimmermann M (1983) Ethical guidelines for investigation of experimental pain in conscious animals. Pain 16:109–110.
30. Ahmed SM, Ahmed S, Tasleem F, Hasan MM, Azhar I. Acute systemic toxicity of four Mimosaceae plants leaves in mice. IOSR J Pharm 2012; 2(2):291-295.
31. Janssen P, Neimemegeers CJE, Dony JGH. The inhibitory effects of Fentanyl and other morphine like analgesics on the warm water induced tail withdrawal reflex in rats. Arzneimittel Forschung 1963; 13: 502–507.
32. Ramabadrnan K, Bansinath M, Turndorf H, Puig MM. Tail immersion test for the evaluation of a nociceptive reaction in mice. Methodological considerations. J Pharmacol Methods 1989; 21(1):21-31.
33. Carter RB. Differentiating analgesic and non-analgesic drug activities on rat hot plate: effect of behavioral endpoint. Pain 1991; 47(2):211-220.
34. Furst S, Gyires K, Knoll J. Analgesic profile of rimazolium as compared to different classes of pain killers. Arzneimittelforschung. 1988; 38(4):552-557.
35. Rujjanawate C, Kanjanapothi D, Panthong A. Pharmacological effect and toxicity of alkaloids from *Gelsemium elegans* Benth. J Ethnopharmacol 2003; 89: 91–95.
36. Asongalem EA, Foyet HS, Ekobo S, Dimo T, Kamtchouing P. Antiinflammatory, lack of central analgesia and antipyretic properties of *Acanthus montanus* (Ness) T. Anderson. J Ethnopharmacol 2004; 95: 63–68.
37. Hosseinzadeh H, Ramezani M, Salmani G. Antinociceptive, anti-inflammatory and acute toxicity effects of *Zataria multiflora* Boiss extracts in mice and rats. J Ethnopharmacol 2003; 73: 379–385.
38. Jinsmaa Y, Okada Y, Tsuda Y, Sasaki Y, Ambo A, Bryant SD, Lazarus LH. Novel 2V, 6Vdimethyl-l-tyrosine-containing pyrazinone opioid mimetic agonists with potent antinociceptive activity in mice. J Pharmacol & Exp Ther 2004; 309: 1–7.
39. Chatu S. Pain management. In: The Hands-on Guide to Clinical Pharmacology. 3rd edition. Wiley-Blackwell Garsington road, Oxford, UK. 2010; p.152.
40. Calixto JB, Beirith A, Ferreira J, Santos ARS, Filho VC, Yunes RA. Naturally occurring antinociceptive substances from plants. Phytother Res 2000;14: 401–418.

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